

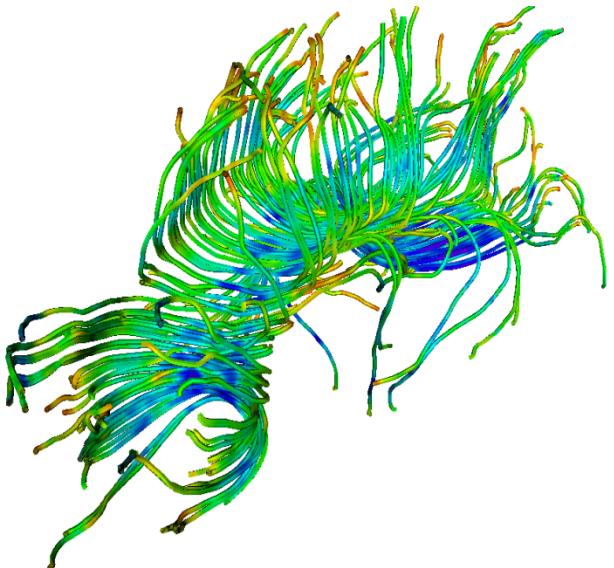


NA-MIC

National Alliance for Medical Image Computing

<http://na-mic.org>

Diffusion Tensor Imaging tutorial

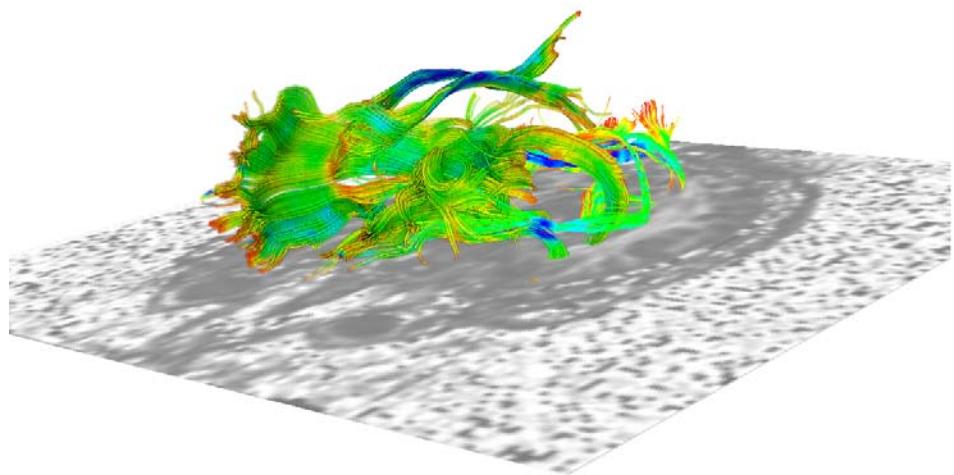


Sonia Pujol, PhD
Surgical Planning Laboratory
Harvard University



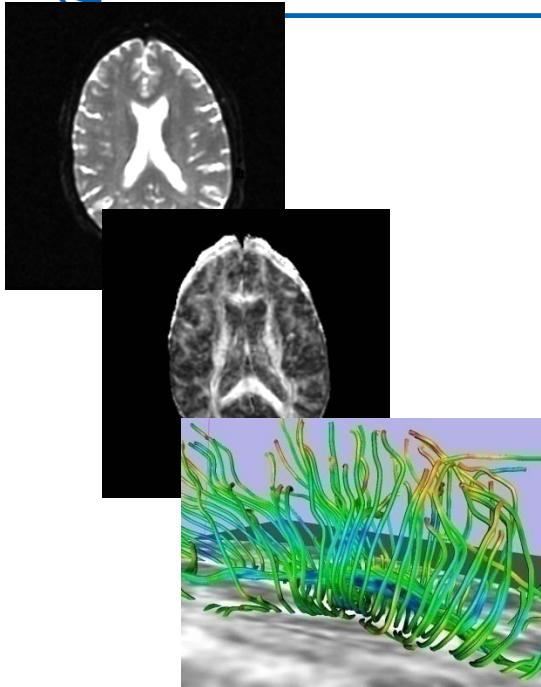
DTI tutorial

This tutorial is an introduction to the advanced **Diffusion MR** capabilities of the **Slicer3** software for medical image analysis.





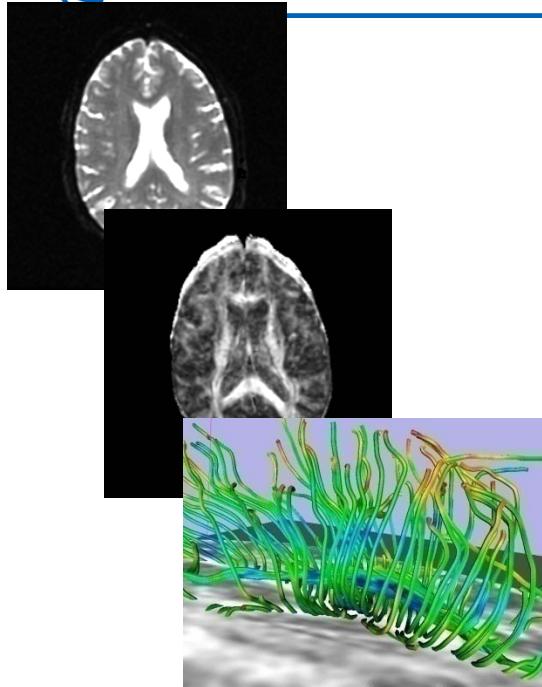
Outline



This tutorial guides you through the process of **loading diffusion MR data**, **estimating diffusion tensors**, and **performing tractography** of white matter bundles.



Outline



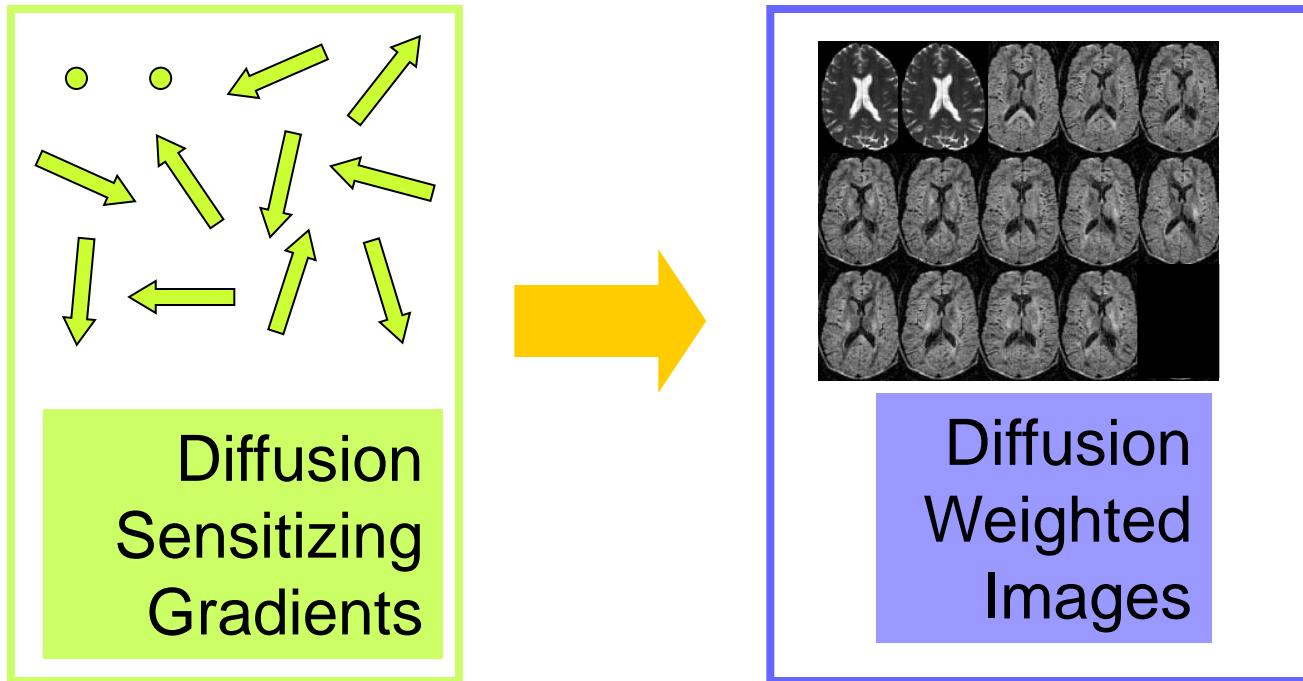
The processing pipeline uses 9 image analysis modules of Slicer3.4

1. Data
2. Volumes
3. Diffusion Tensor Estimation
4. Diffusion Tensor Scalar Measurements
5. Editor
6. LabelMap Seeding
7. Fiber Bundles
8. Fiducials
9. Fiducial Seeding



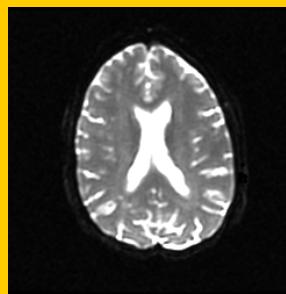
Tutorial Dataset

The Diffusion MR tutorial dataset is composed of a **Diffusion Weighted MR scan** of the brain acquired with 12 gradient directions and 2 baseline.

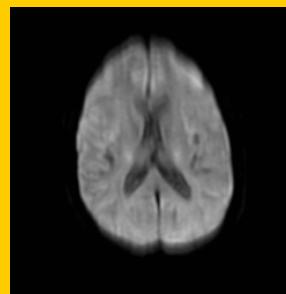




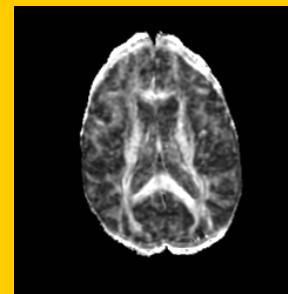
DTI Processing Pipeline



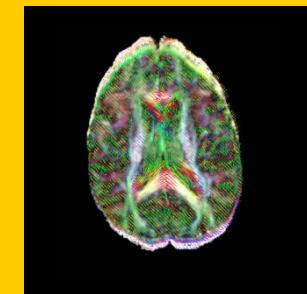
DWI
Acquisition



Tensor
Calculation



Scalar
Maps



3D
Visualization



Start Slicer3

Linux/Mac users

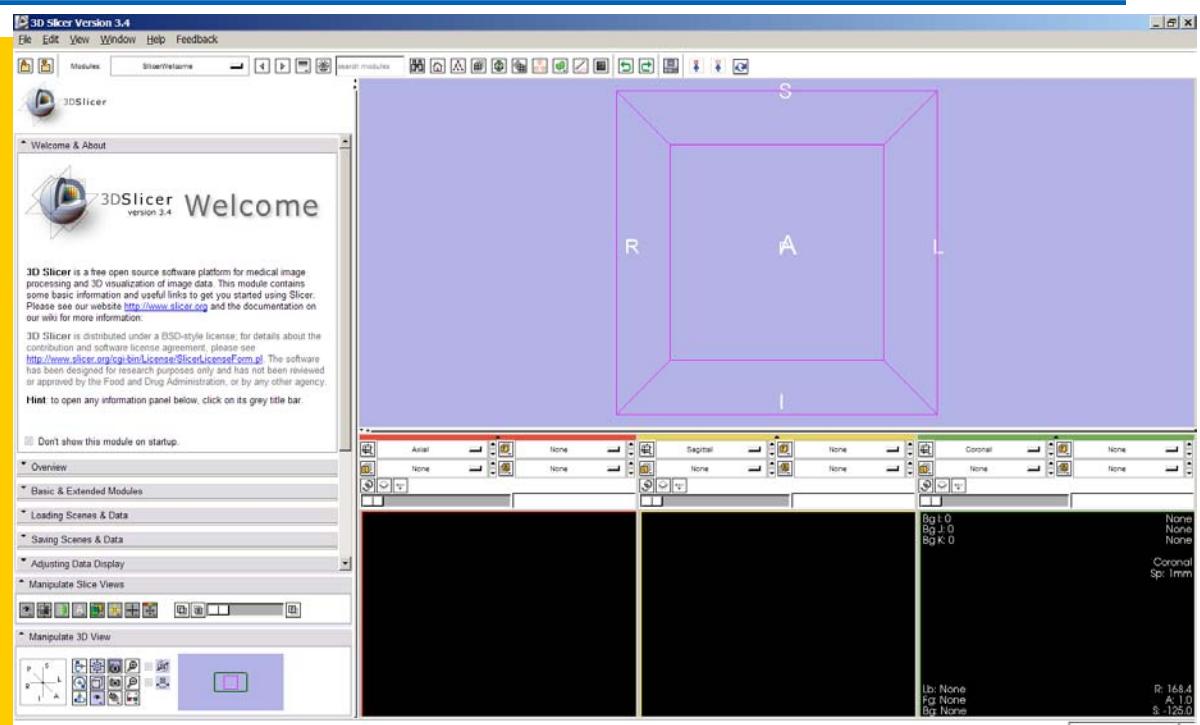
Launch the Slicer3 executable located in the Slicer3.4 directory

Windows users

Select

Start → All Programs

→ Slicer3 3.4 2009-05-21 → Slicer3



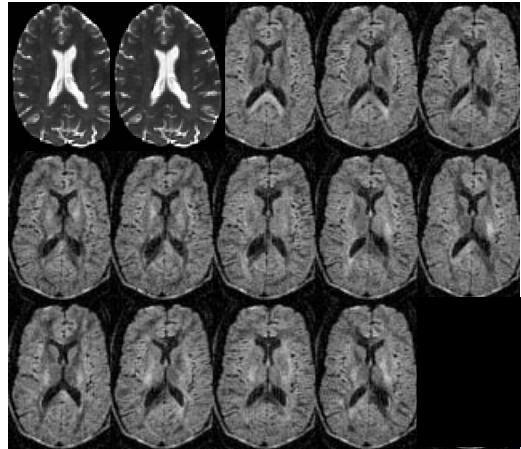


Slicer Welcome

The screenshot shows the Slicer3 application window. A yellow box highlights the left sidebar where the 'SlicerWelcome' module is selected. The main area displays a 3D volume rendering of a brain with a coordinate system (Sagittal, Coronal, Axial) and some text labels. The bottom right corner of the main window shows slice parameters: R: 168.4, A: 1.0, S: -125.0.

The **SlicerWelcome** module is the module displayed by default.

This module gives an overview of the GUI of Slicer3, and data loading & saving functionalities.



$$S_i = S_0 e^{-b\hat{g}^T \underline{D} \hat{g}_i}$$

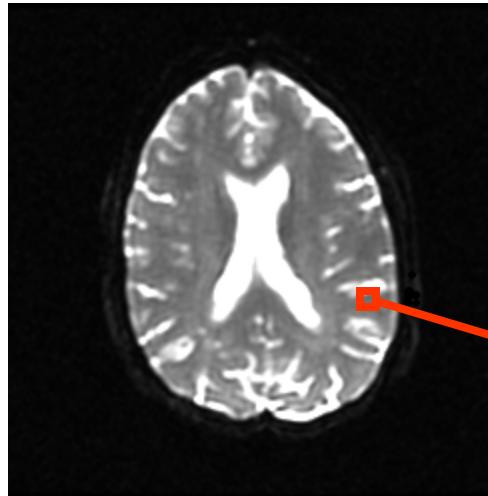
Part 1:

Diffusion data loading and tensor estimation



Diffusion Tensor

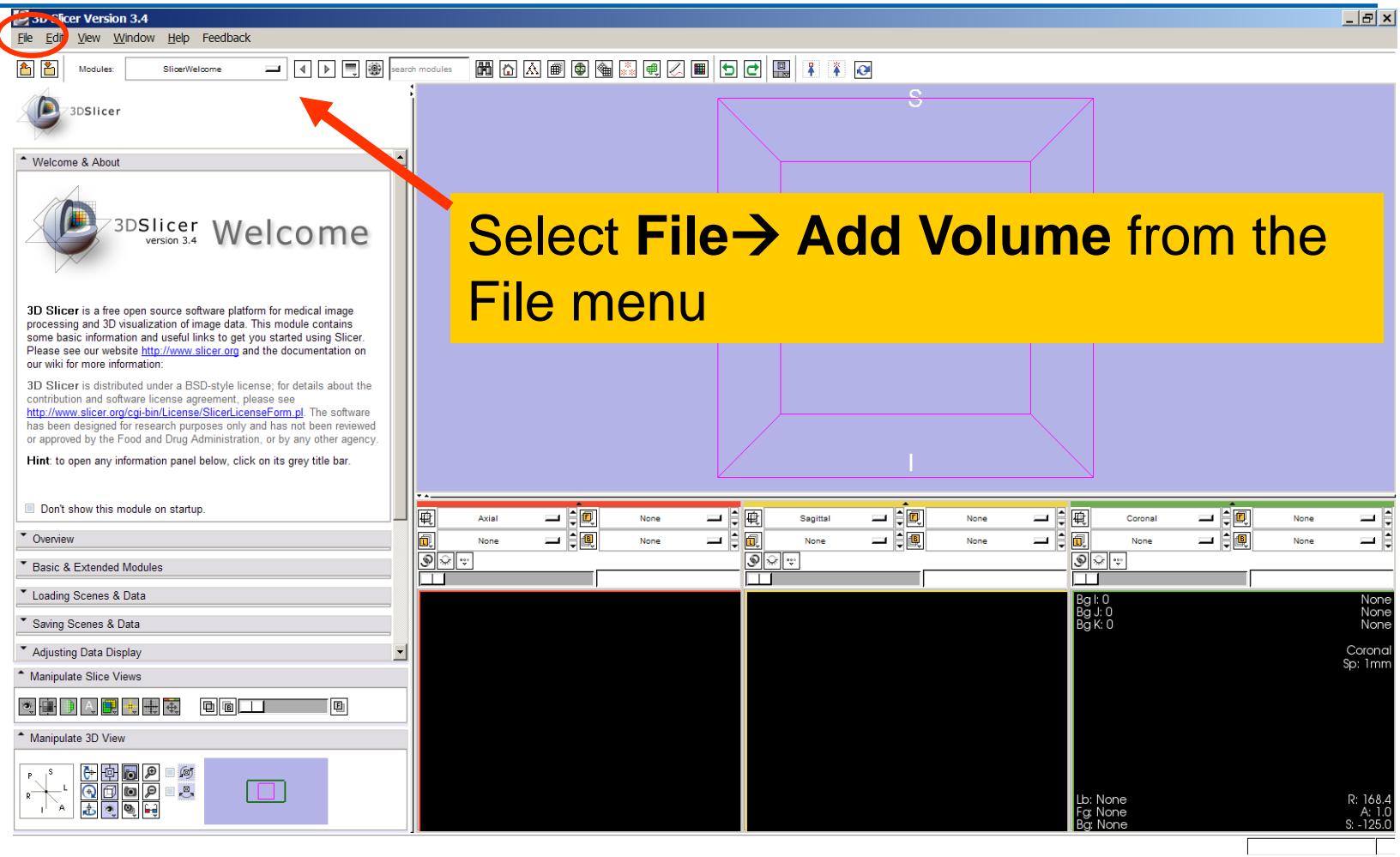
Stejskal-Tanner



$$\underline{D} = \begin{bmatrix} D_{xx} & D_{xy} & D_{xz} \\ D_{yx} & D_{yy} & D_{yz} \\ D_{zx} & D_{zy} & D_{zz} \end{bmatrix}$$

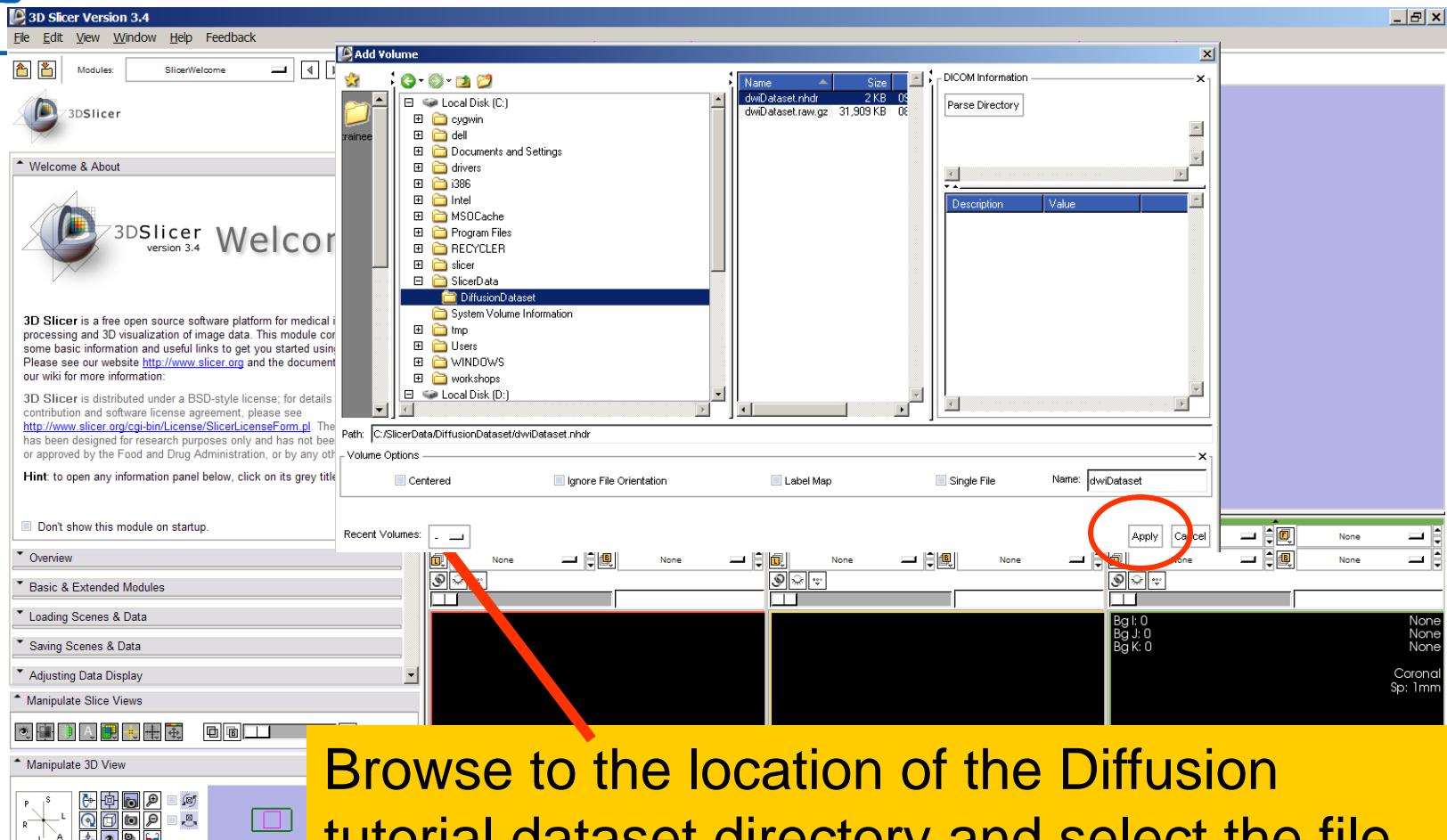


Loading the DWI volume





Loading the DWI volume



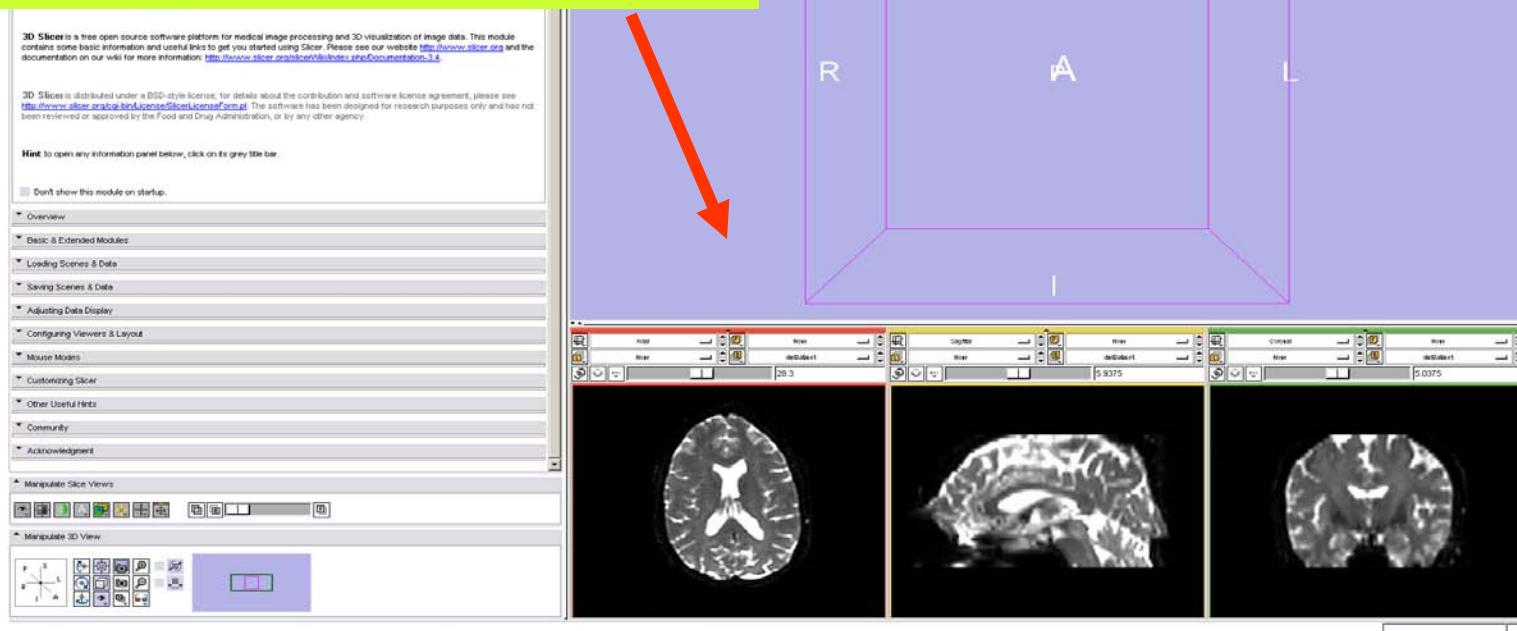
Browse to the location of the Diffusion tutorial dataset directory and select the file **dwiDataset.nhdr**

Click on **Apply** to load the volume



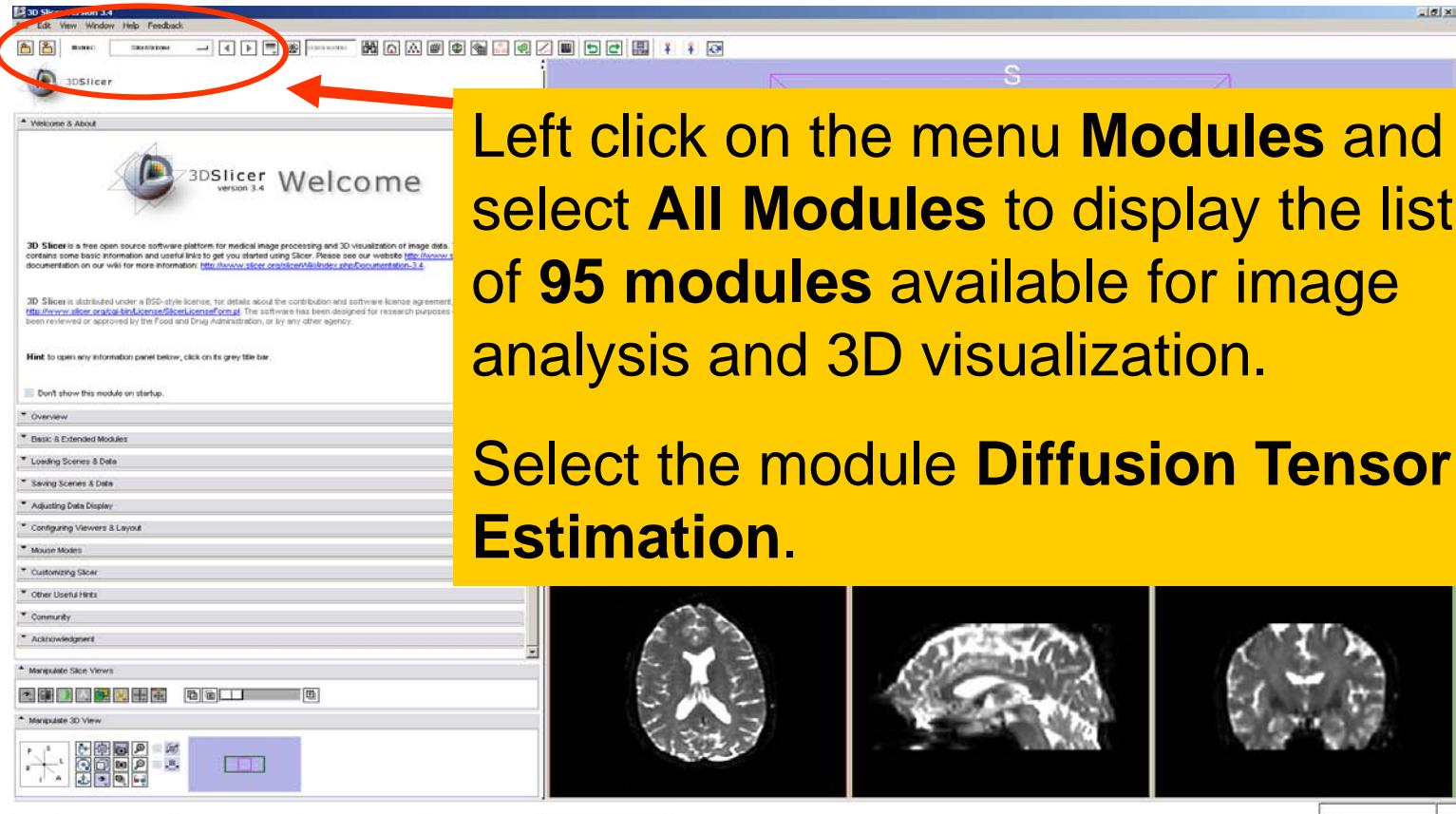
Loading the DWI volume

Slicer displays the anatomical views of the baseline volume of the diffusion dataset in the 2D Slice Viewer.



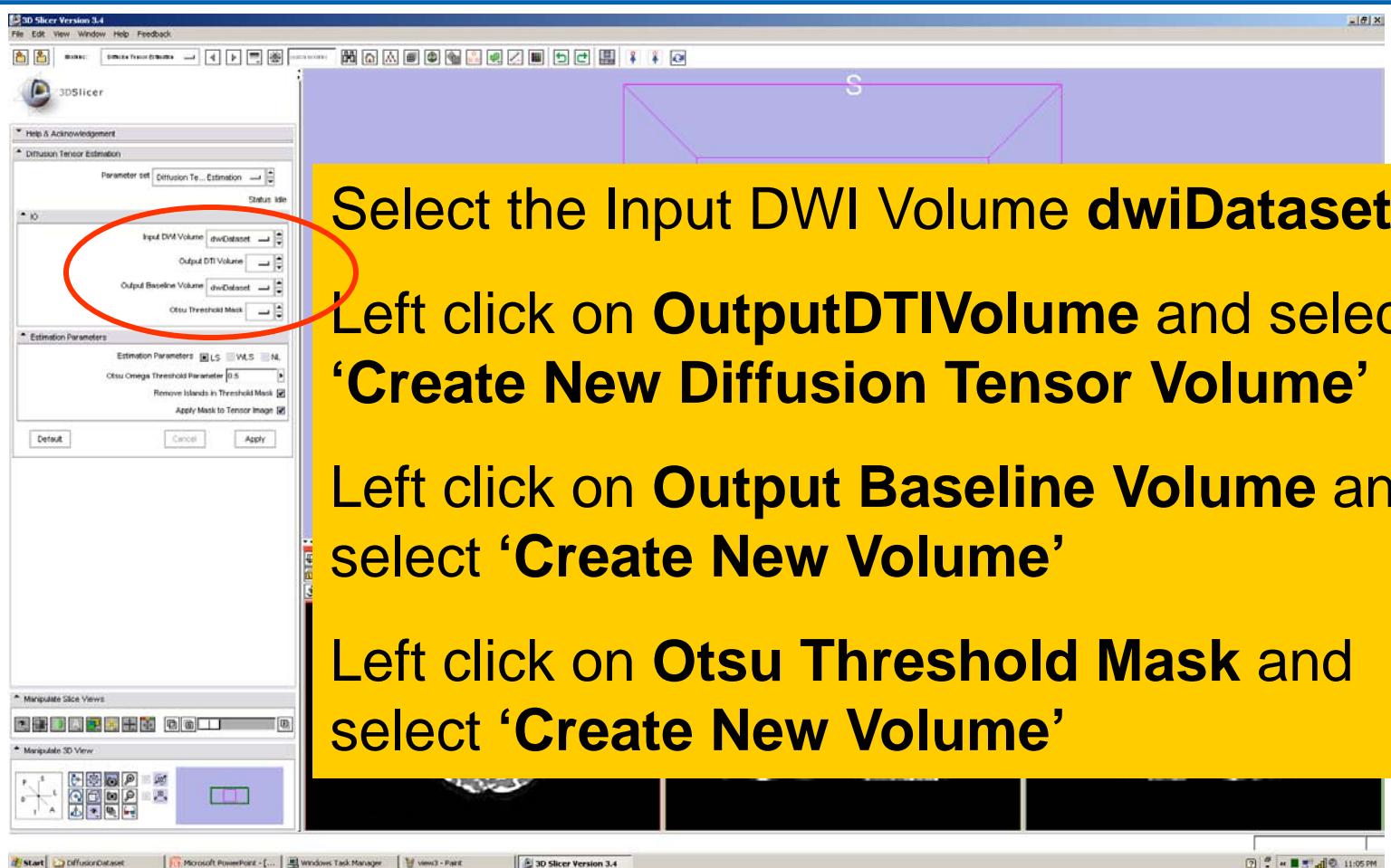


Tensor Estimation



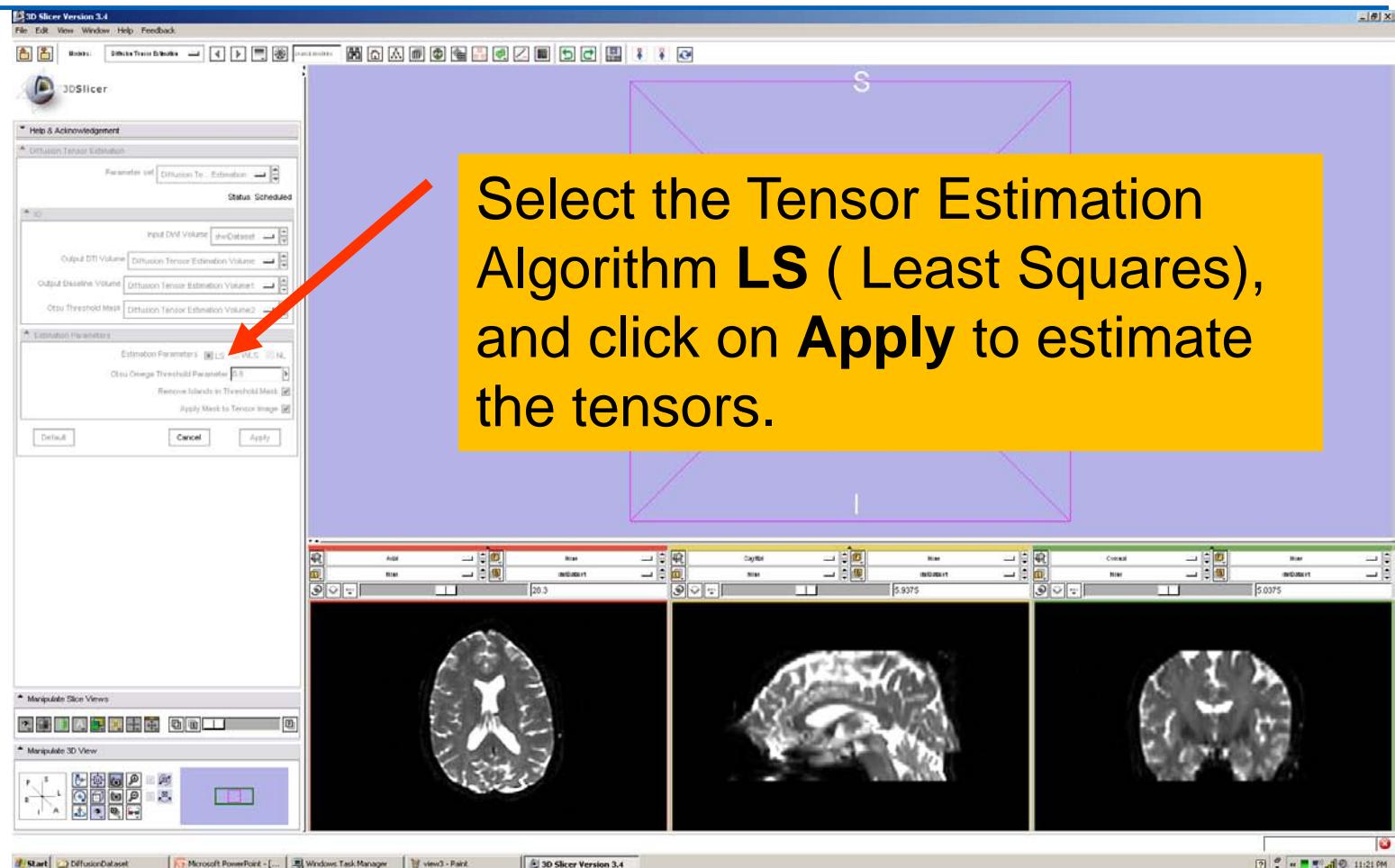


Tensor Estimation



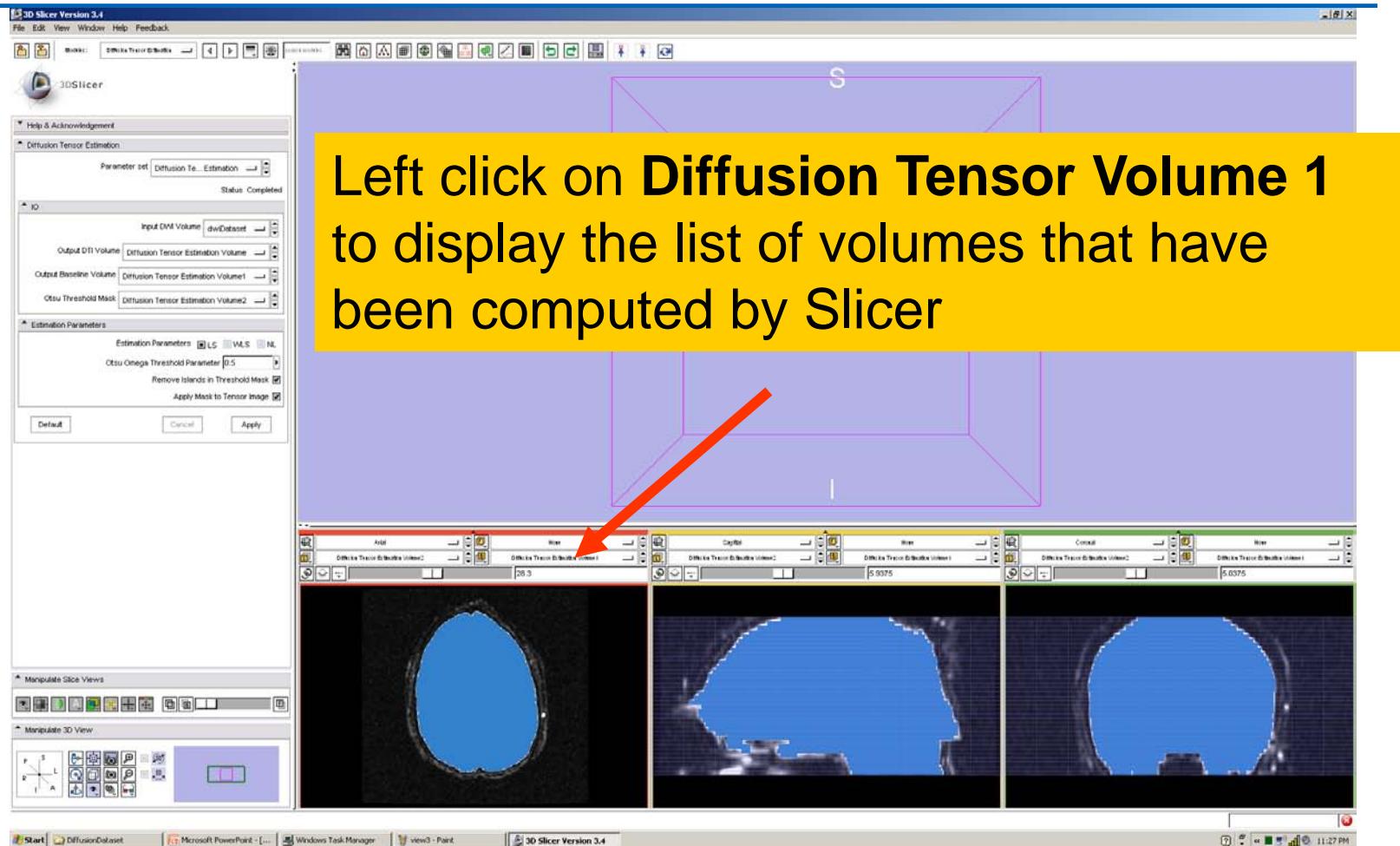


Tensor Estimation





Tensor Estimation





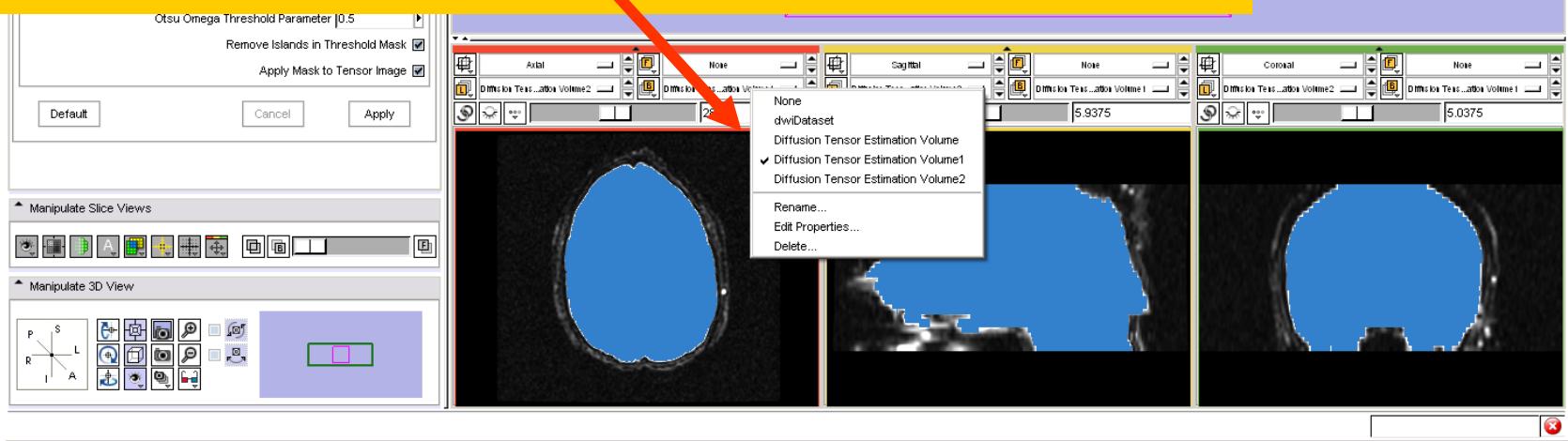
Tensor Estimation

3D Slicer Version 3.4

Diffusion Tensor Estimation Volume is the volume of estimated tensors

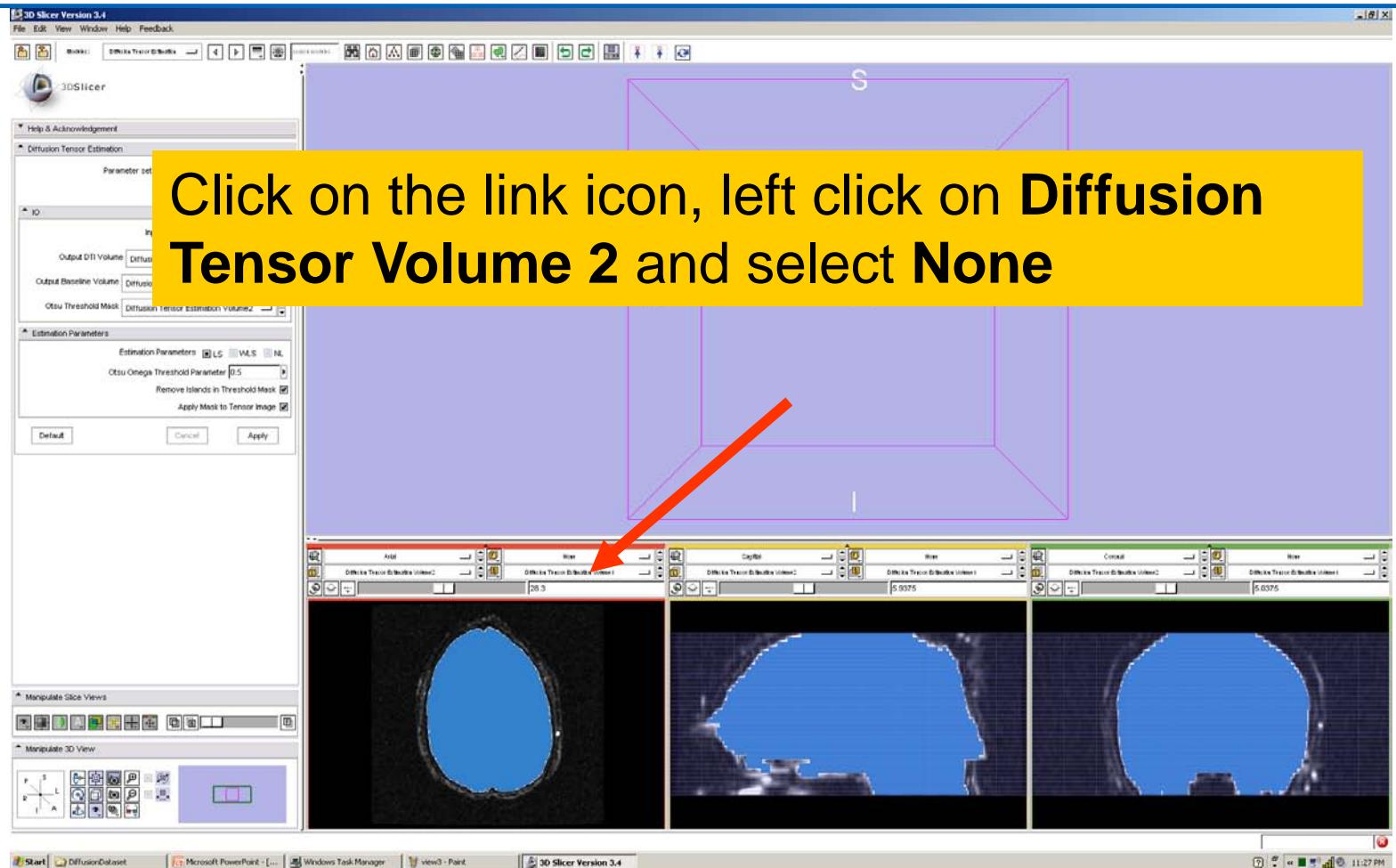
Diffusion Tensor Estimation Volume 1 is the Baseline volume

Diffusion Tensor Estimation Volume 2 is the tensor mask (blue)



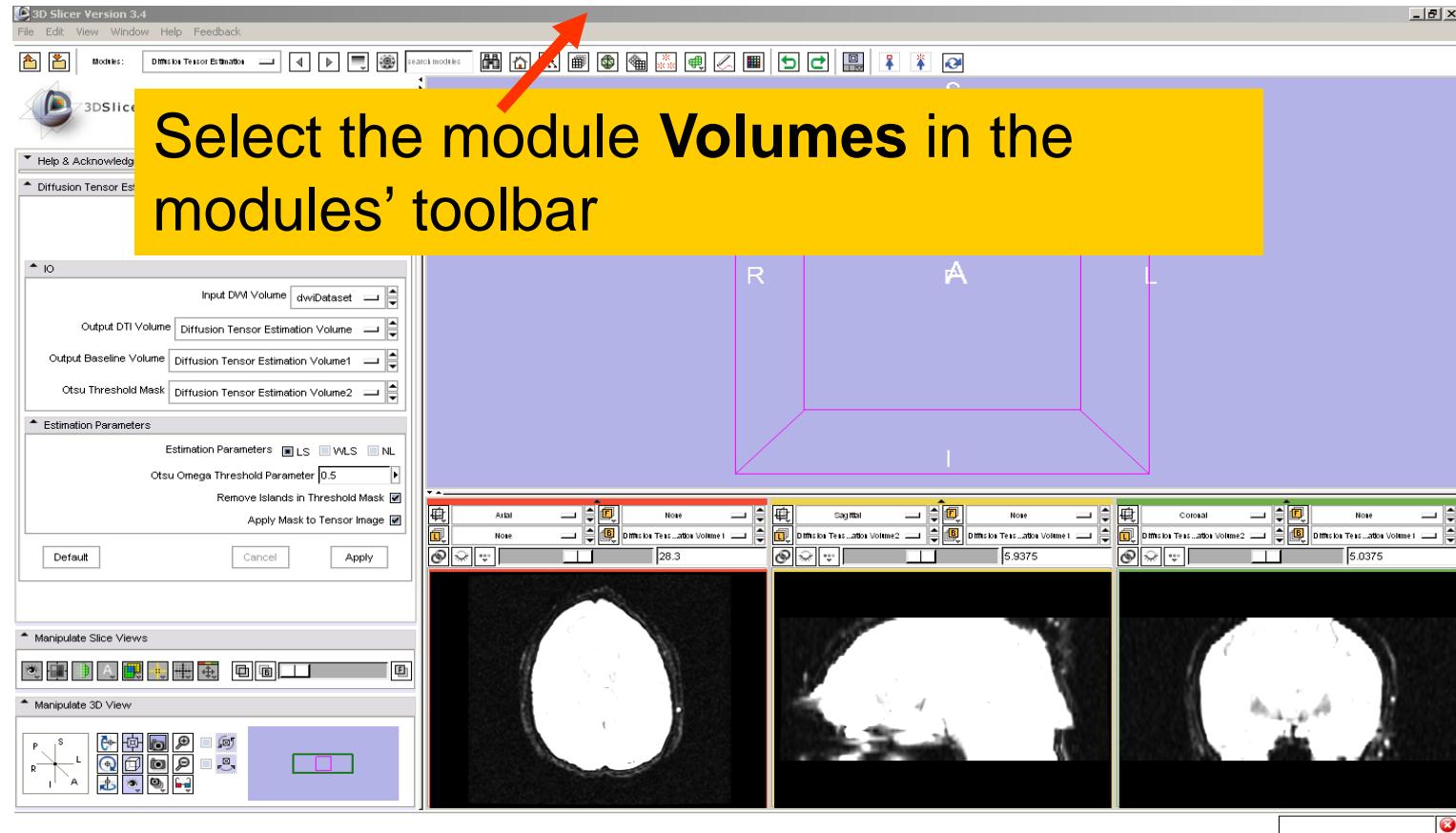


Tensor Estimation





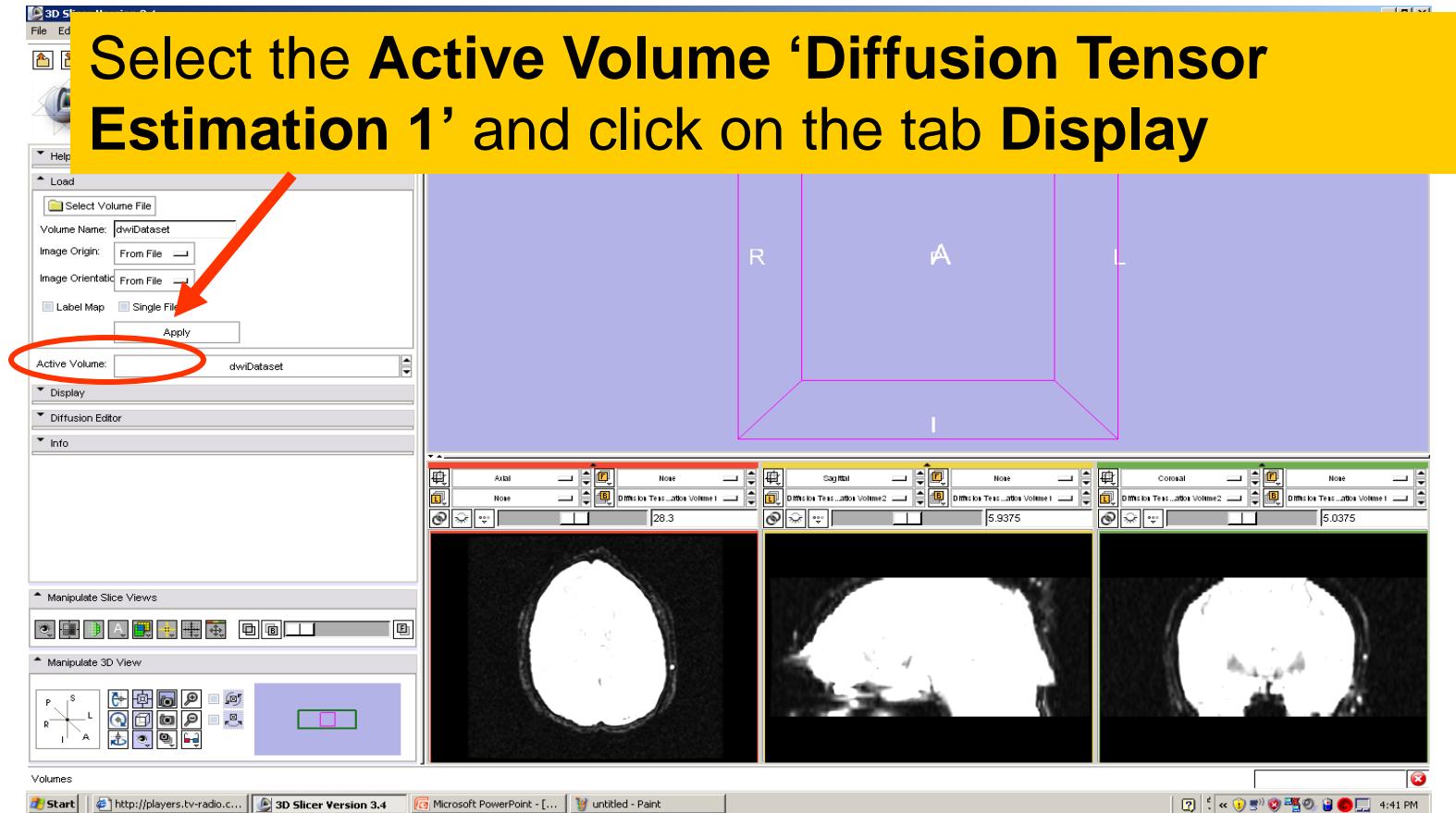
Tensor Estimation





Tensor Estimation

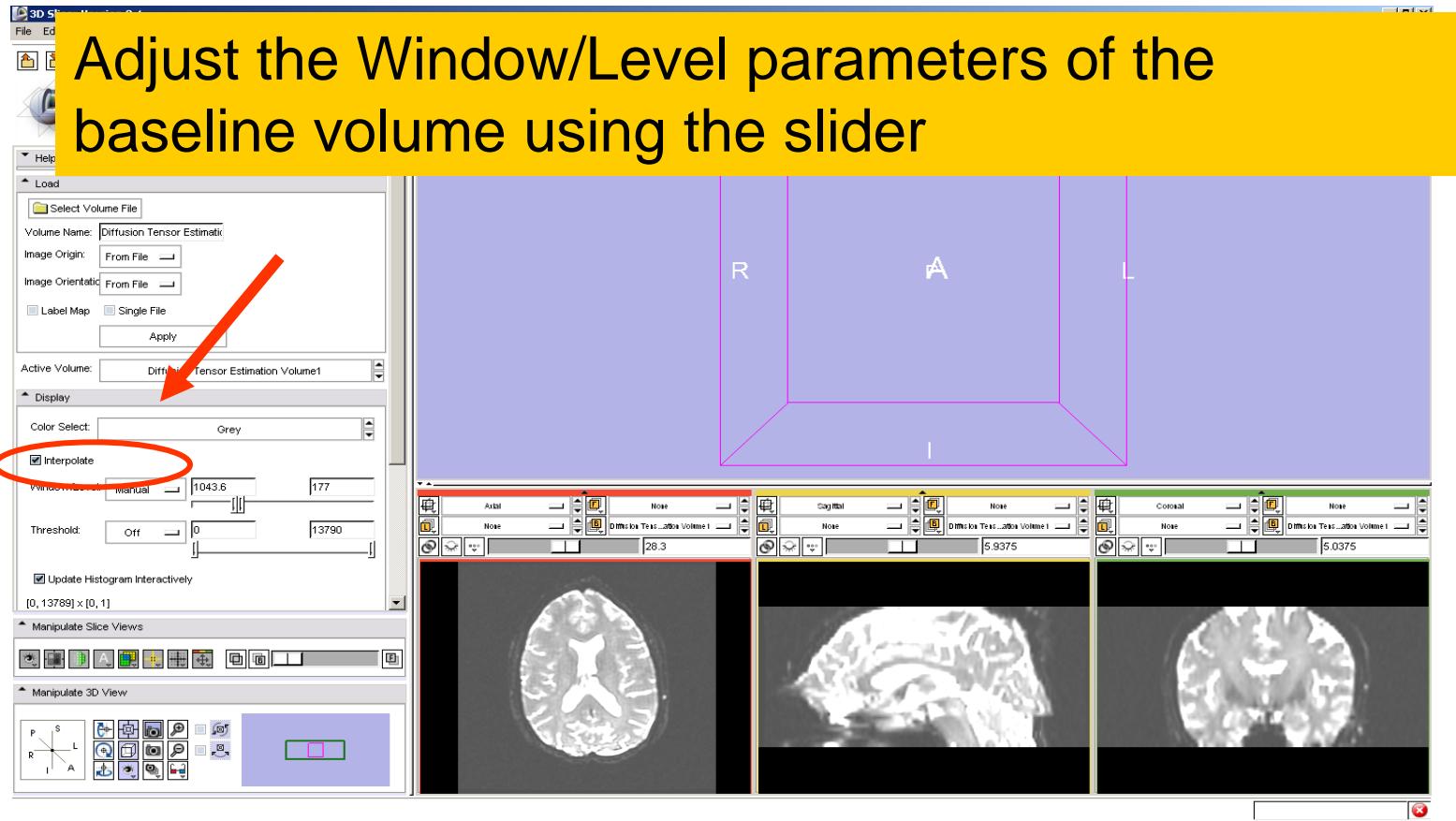
Select the Active Volume 'Diffusion Tensor Estimation 1' and click on the tab **Display**





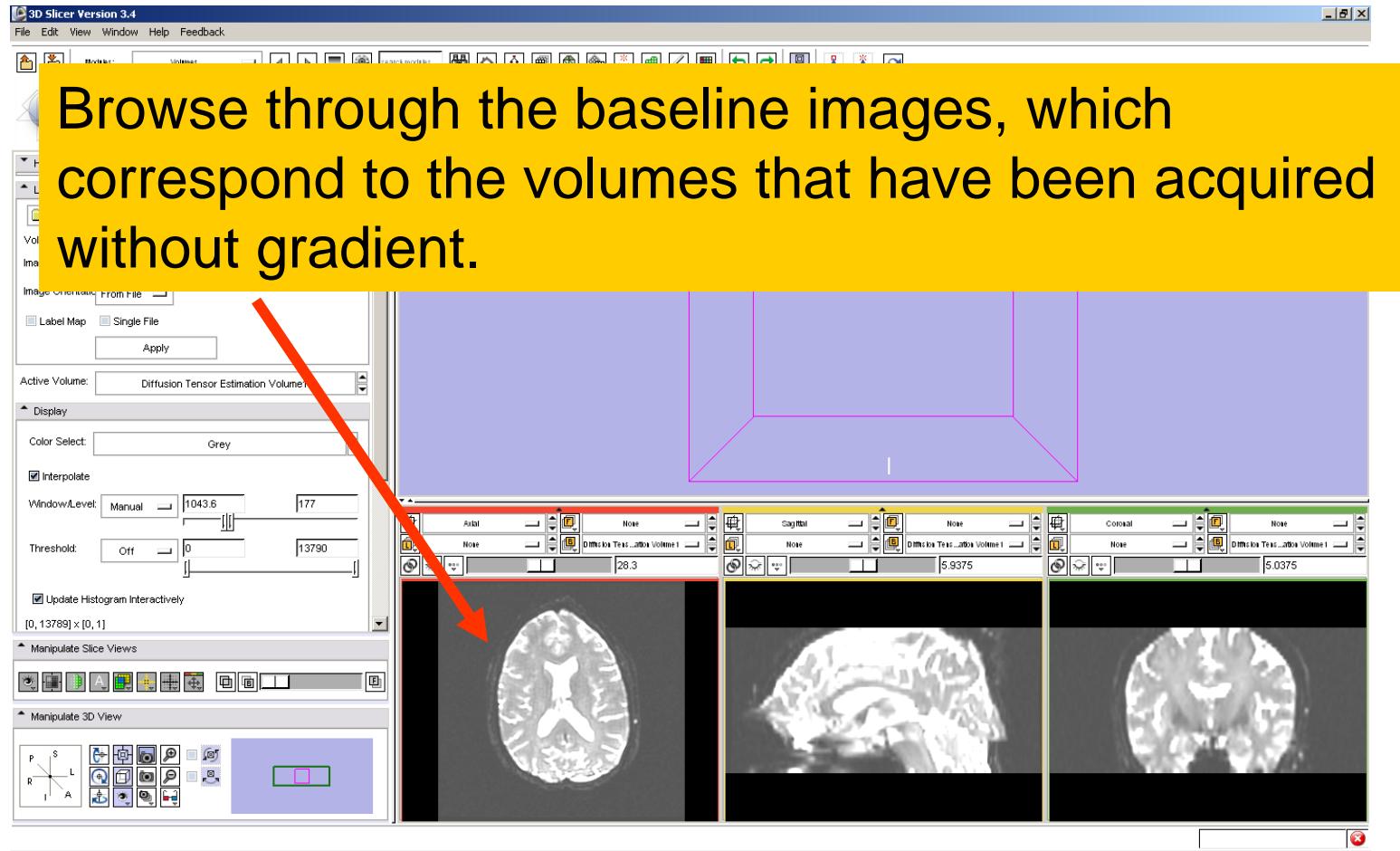
Tensor Estimation

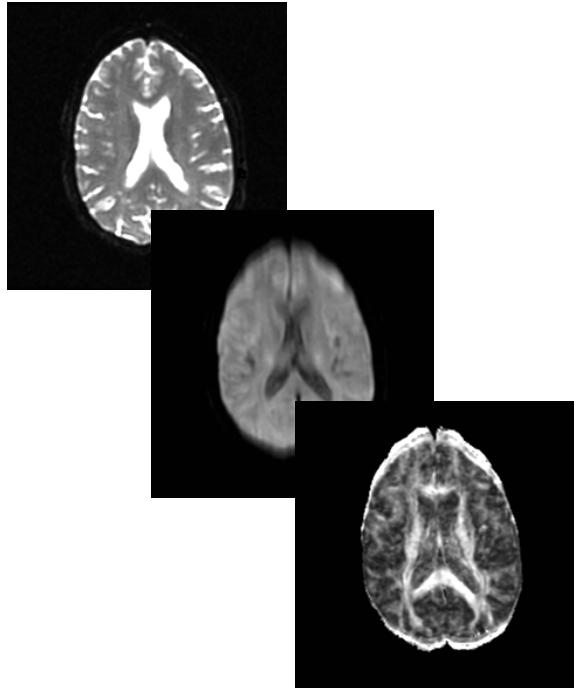
Adjust the Window/Level parameters of the baseline volume using the slider





Tensor Estimation



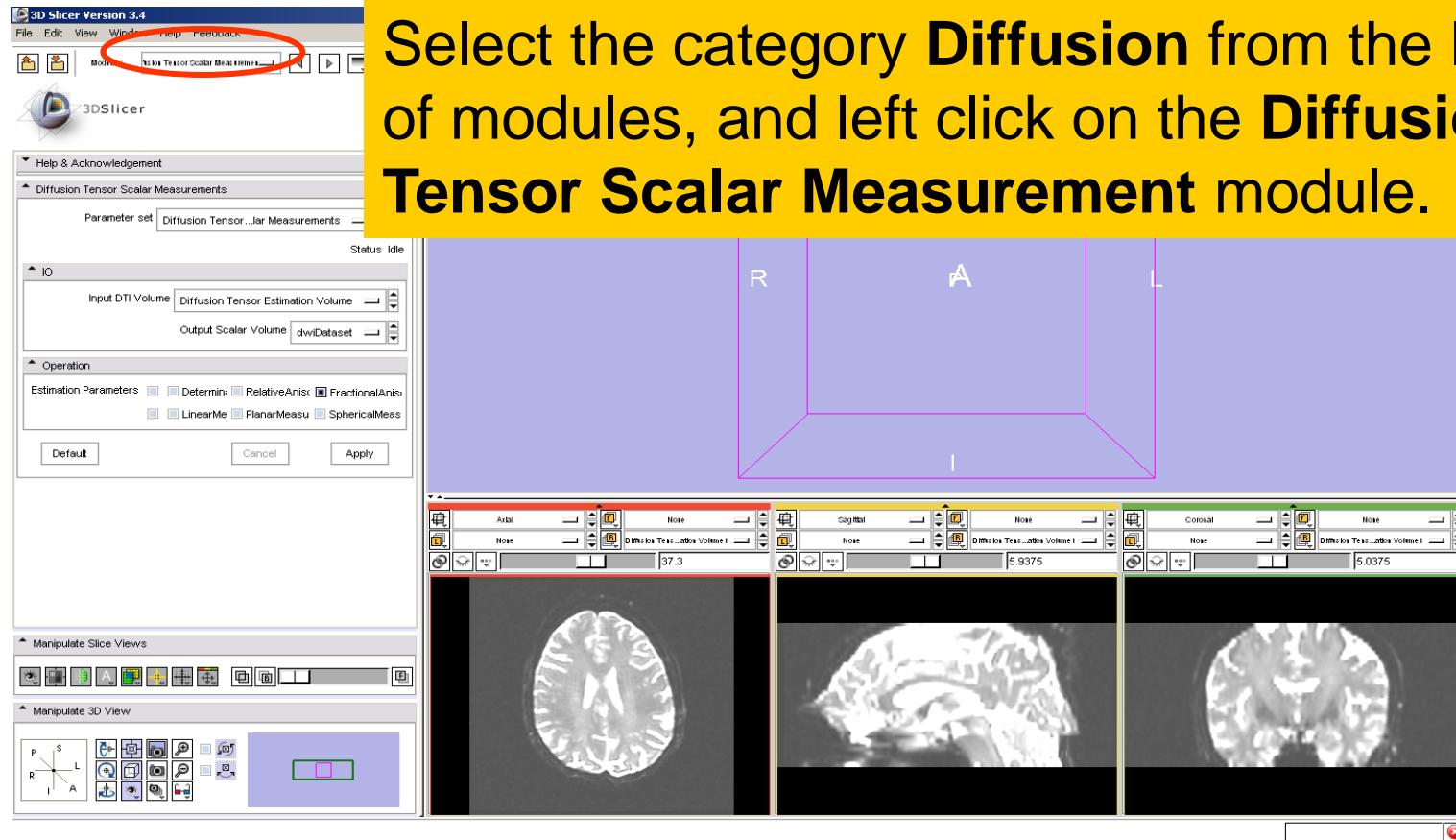


Part2:

Scalar Measurements

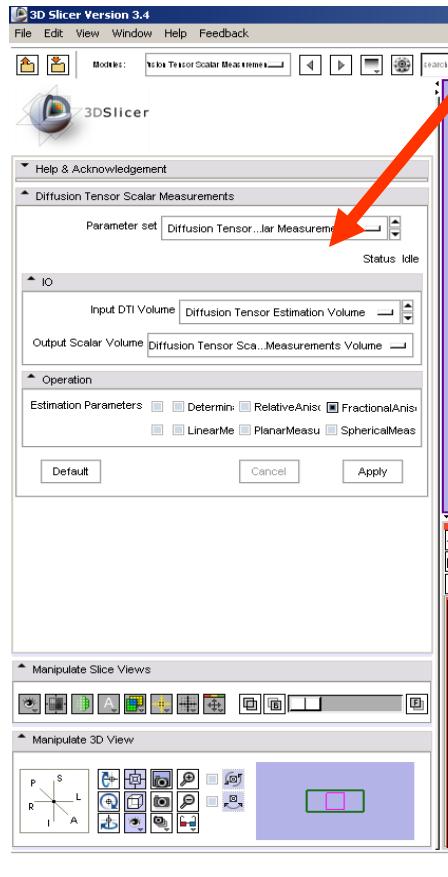


Scalar Measurements





Scalar Measurements



Select the Input DTI Volume **Diffusion Tensor Estimation Volume**

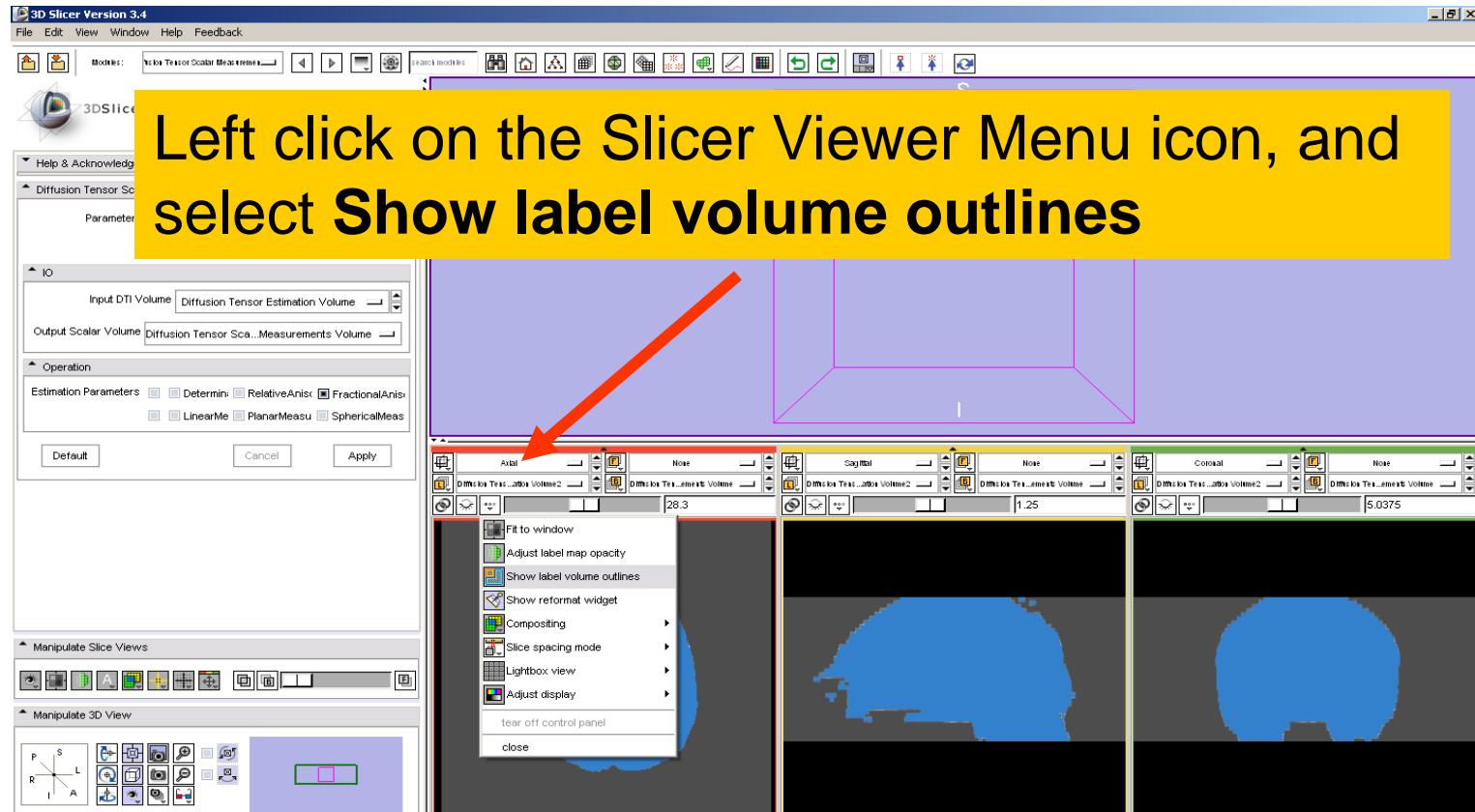
Select the Output Scalar Volume
'Create New Volume'

Select the Operation **Fractional Anisotropy**, and click on **Apply**



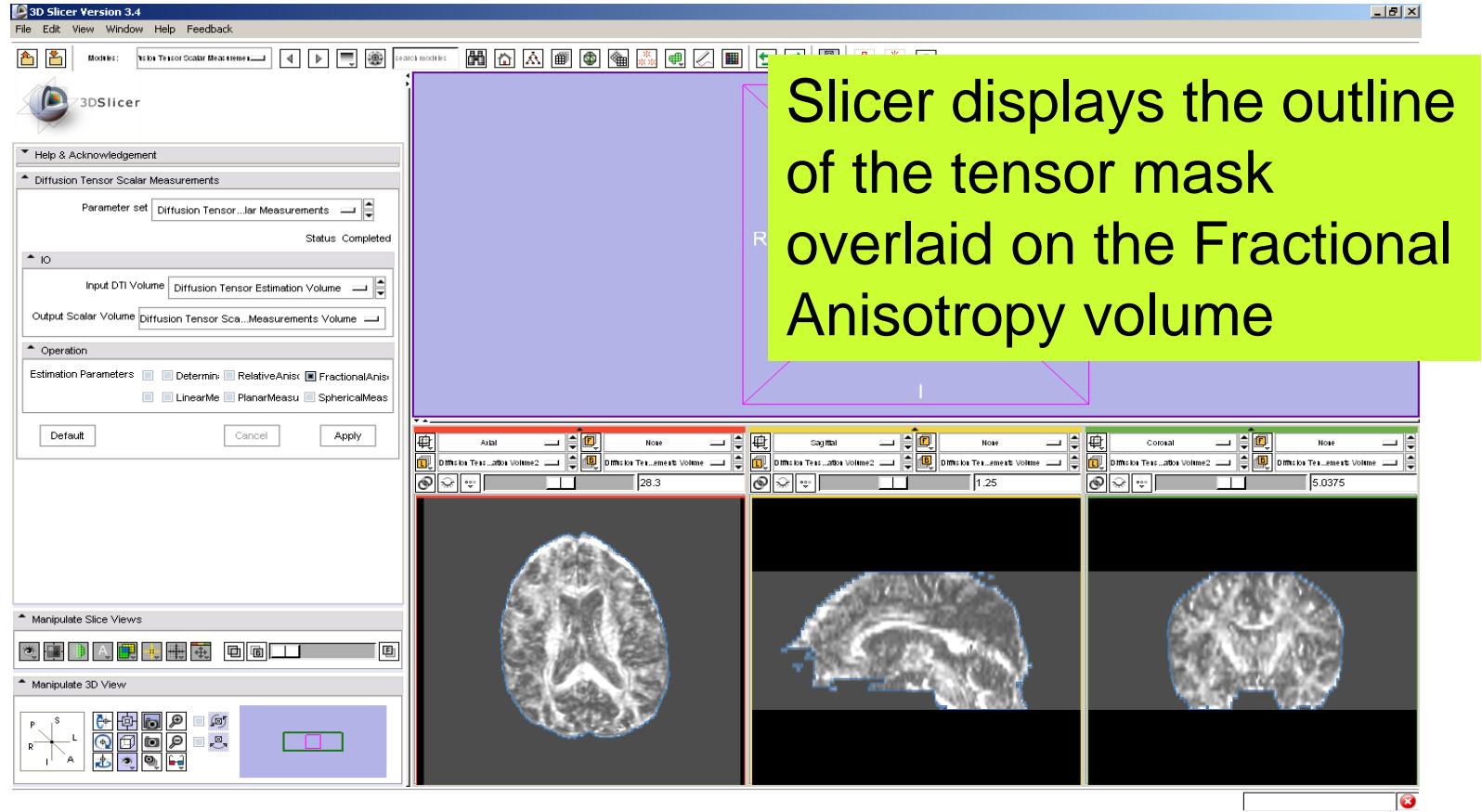


Fractional Anisotropy Volume



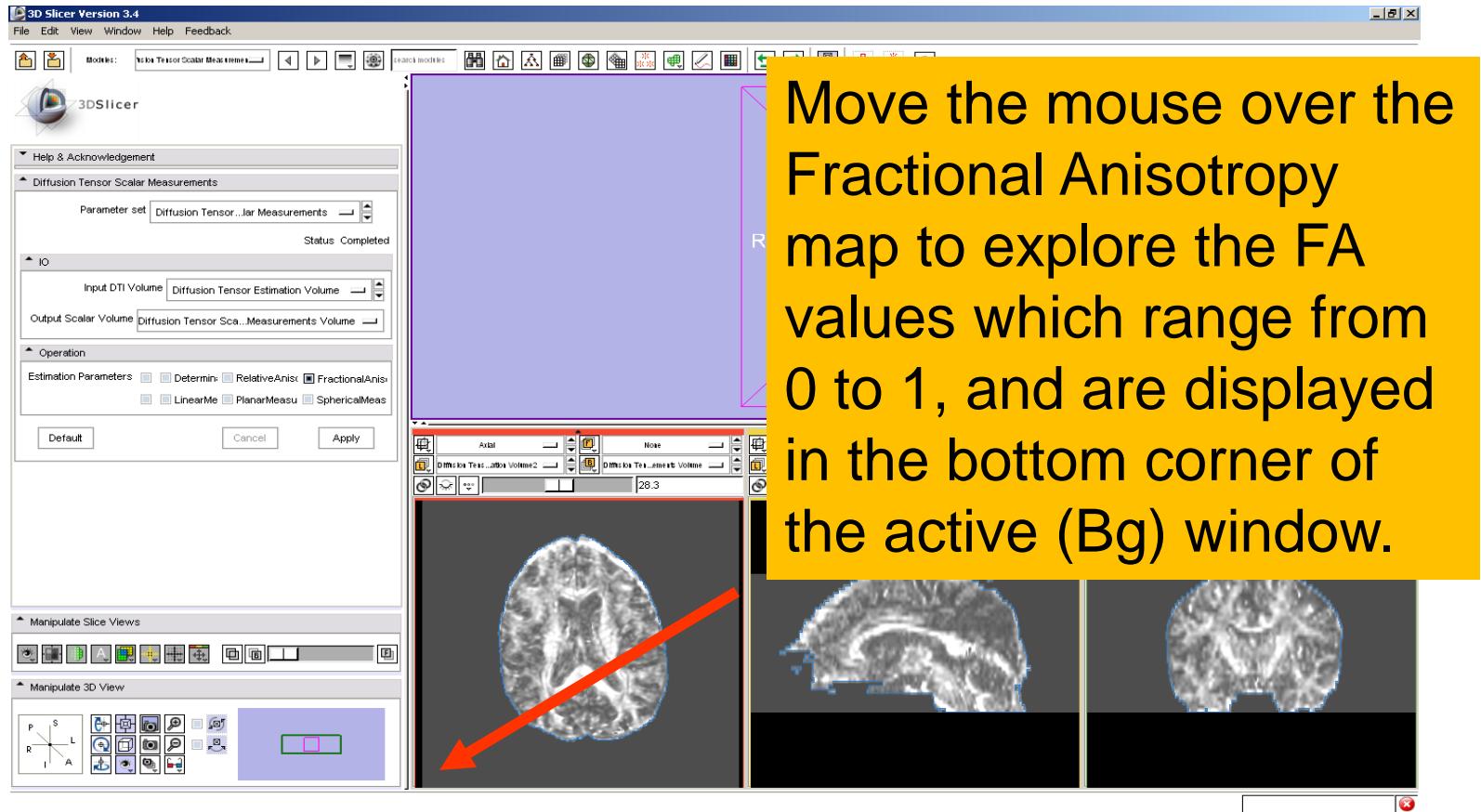


Fractional Anisotropy Volume





Fractional Anisotropy Volume



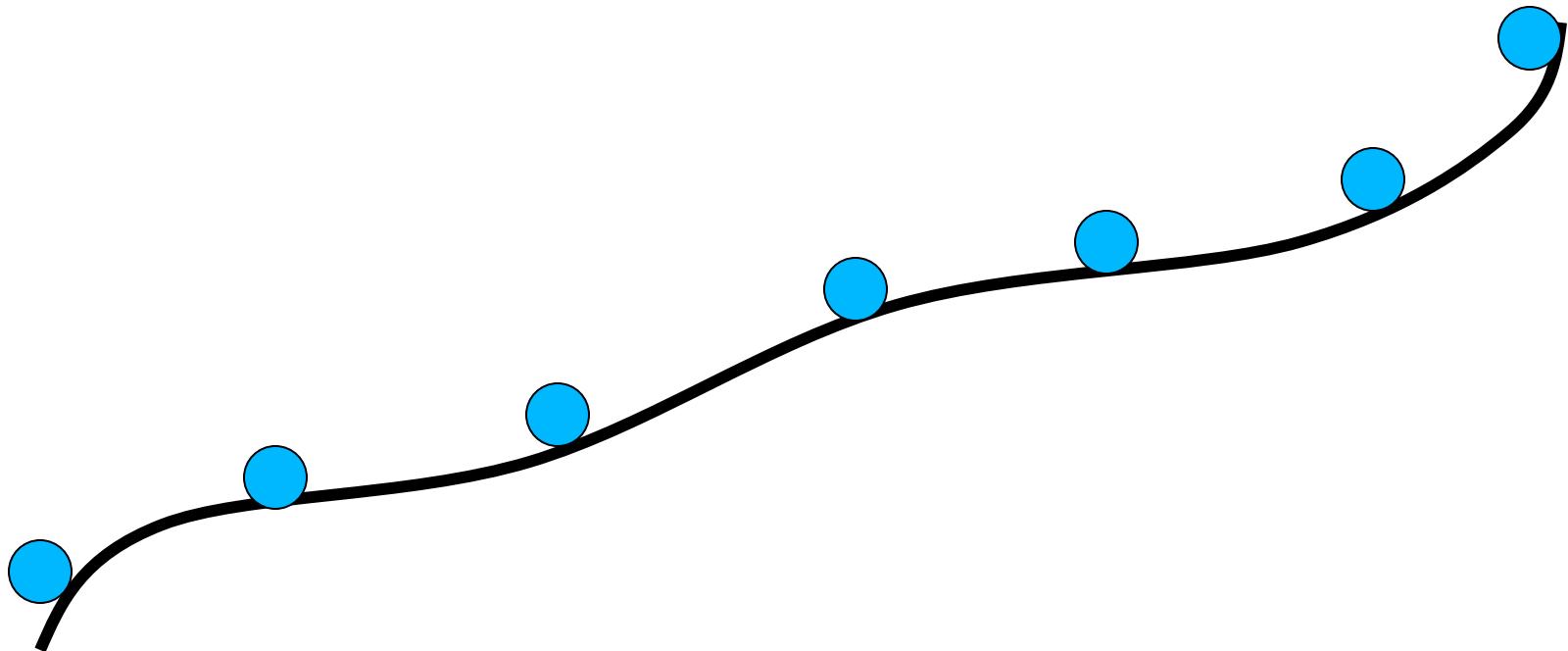


Part 3:

Region of Interest based Tractography

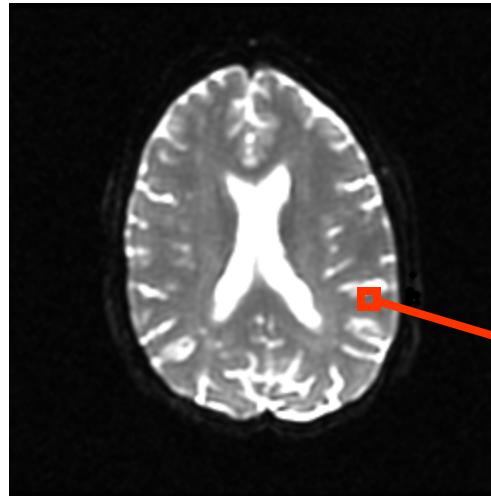


Tractography





Diffusion Tensor



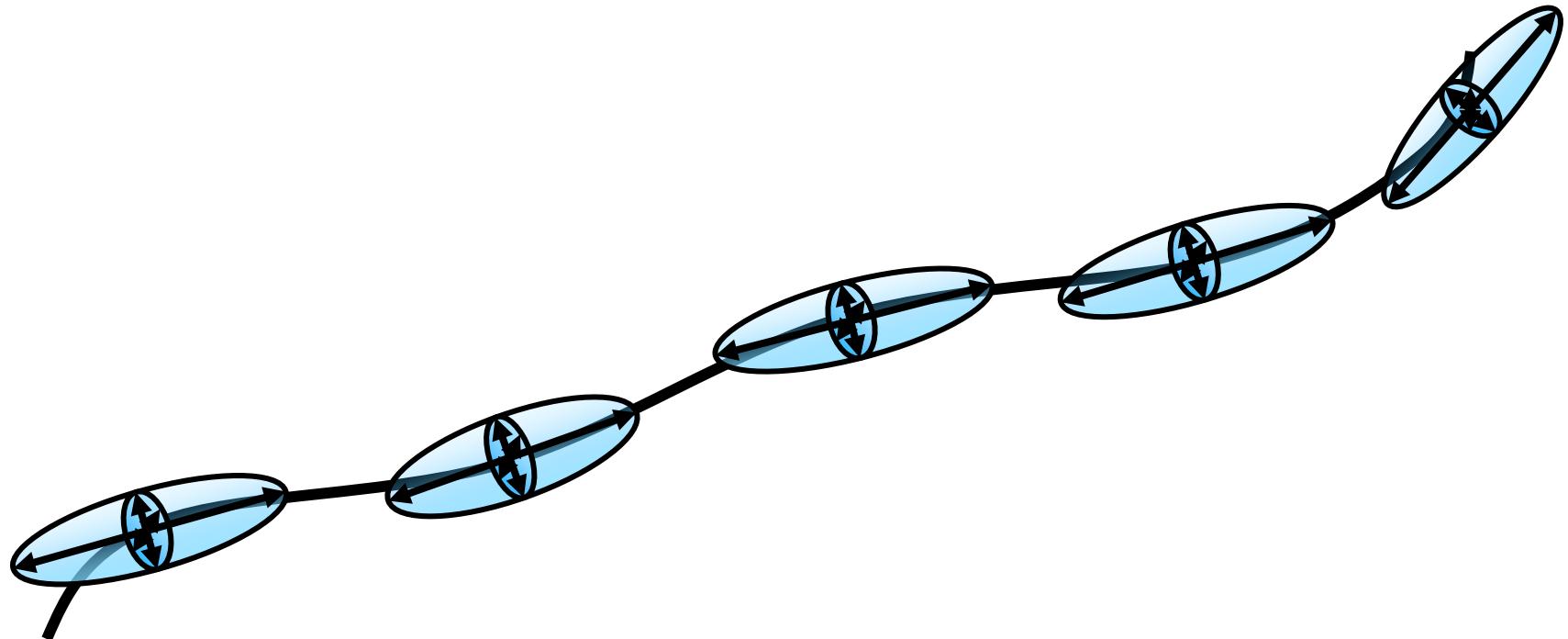
Stejskal-Tanner

$$S_i = S_0 e^{-b \hat{g} i^T \underline{D} \hat{g}_i}$$

$$\underline{D} = \begin{bmatrix} D_{xx} & D_{xy} & D_{xz} \\ D_{yx} & D_{yy} & D_{yz} \\ D_{zx} & D_{zy} & D_{zz} \end{bmatrix}$$

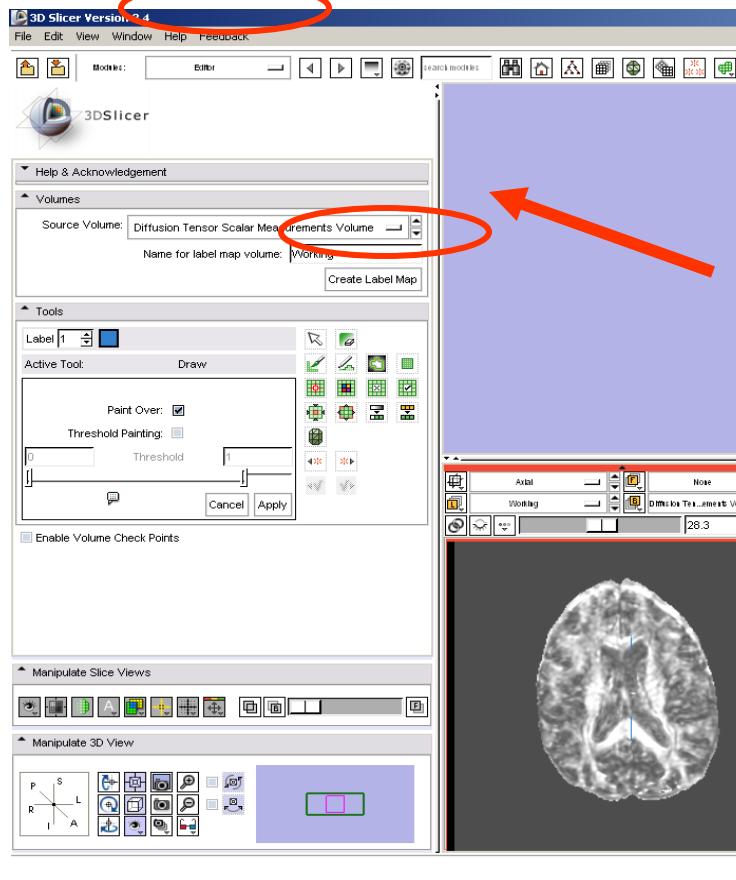


Tractography





LabelMap Generation



Select the module Editor
in the modules' menu.



Select the Source Volume

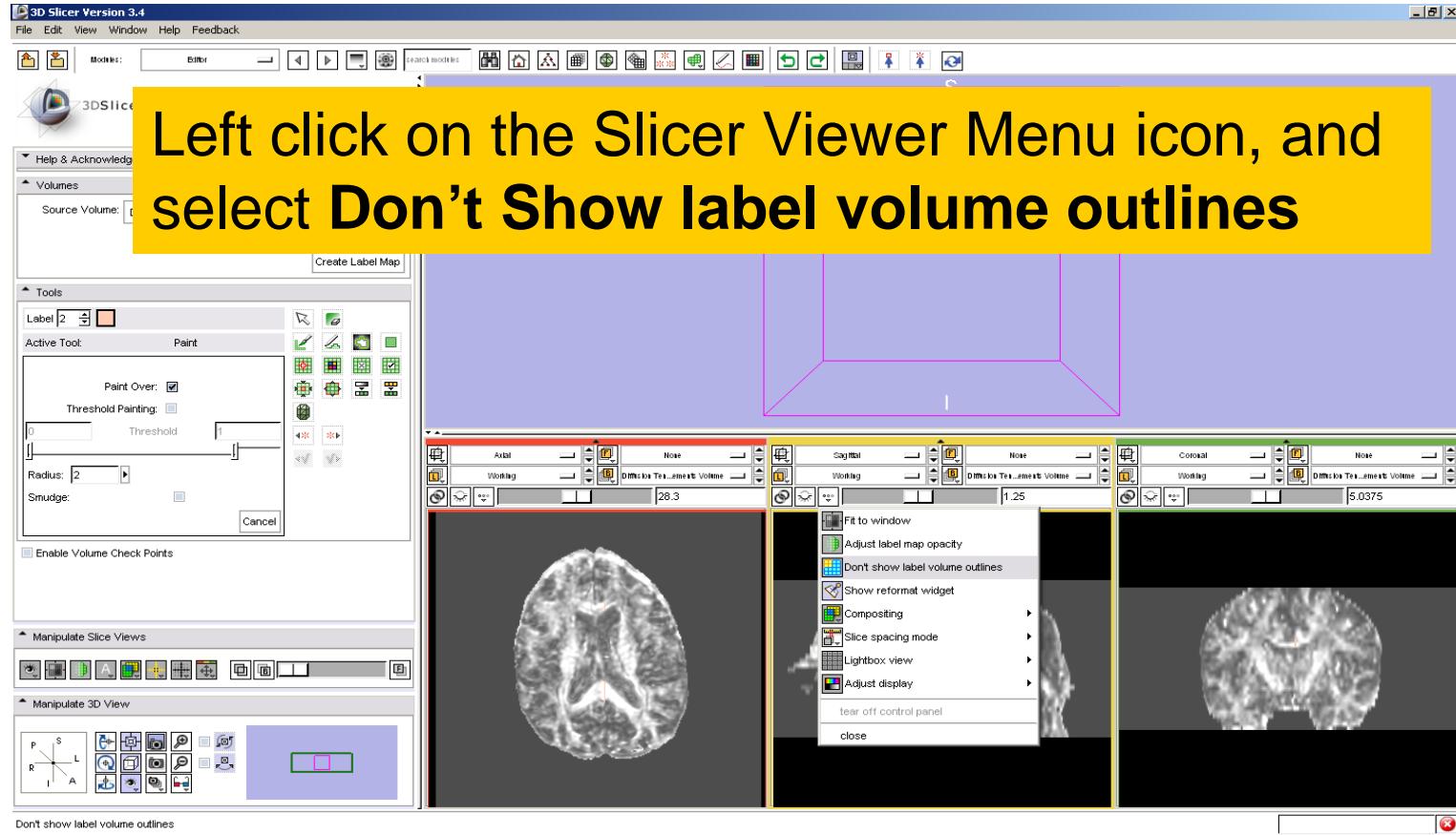
**Diffusion Tensor Scalar
Measurements Volume**

Select the label map
volume **Working** and click
on **Create Label Map**



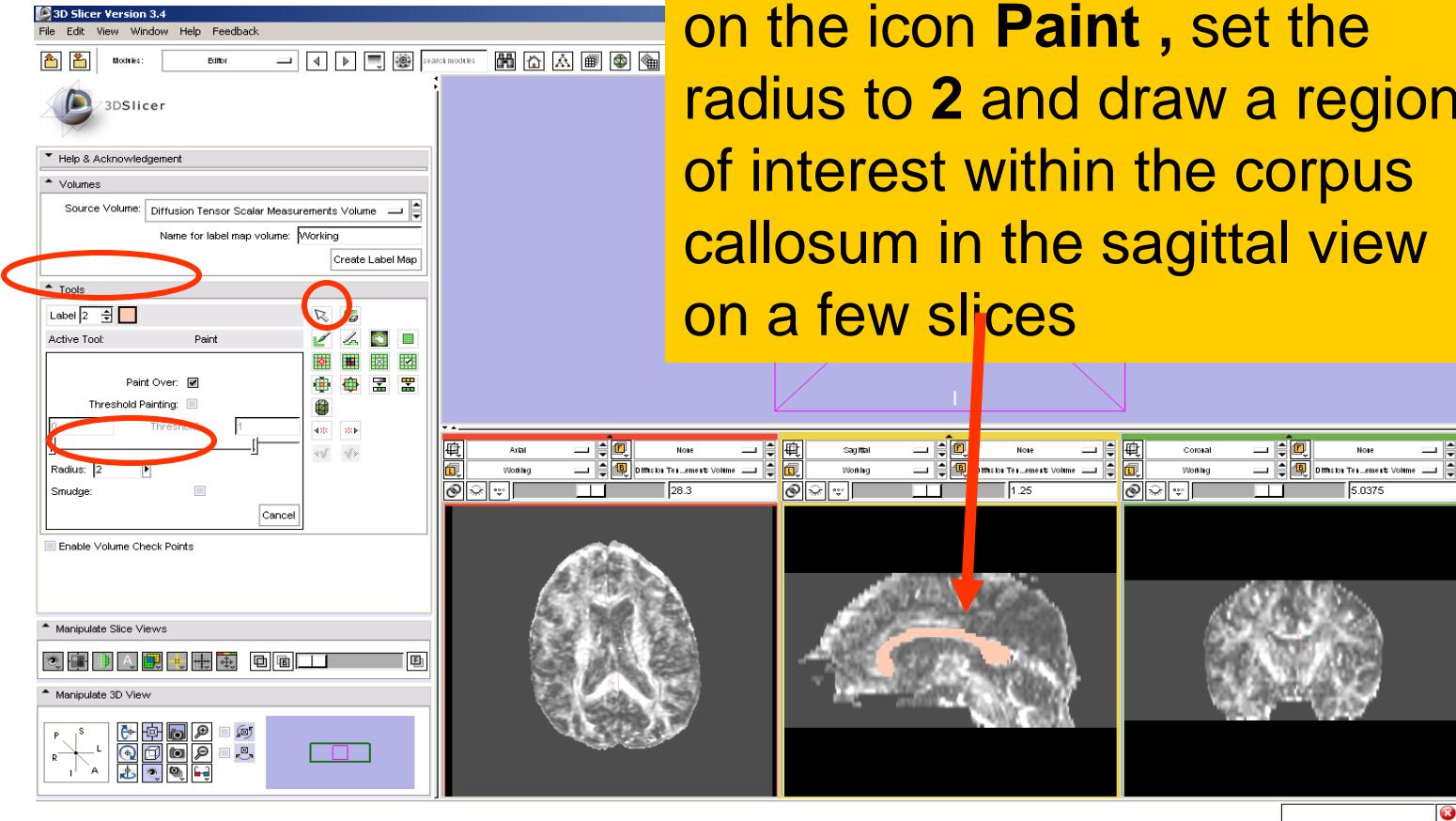


LabelMap Generation



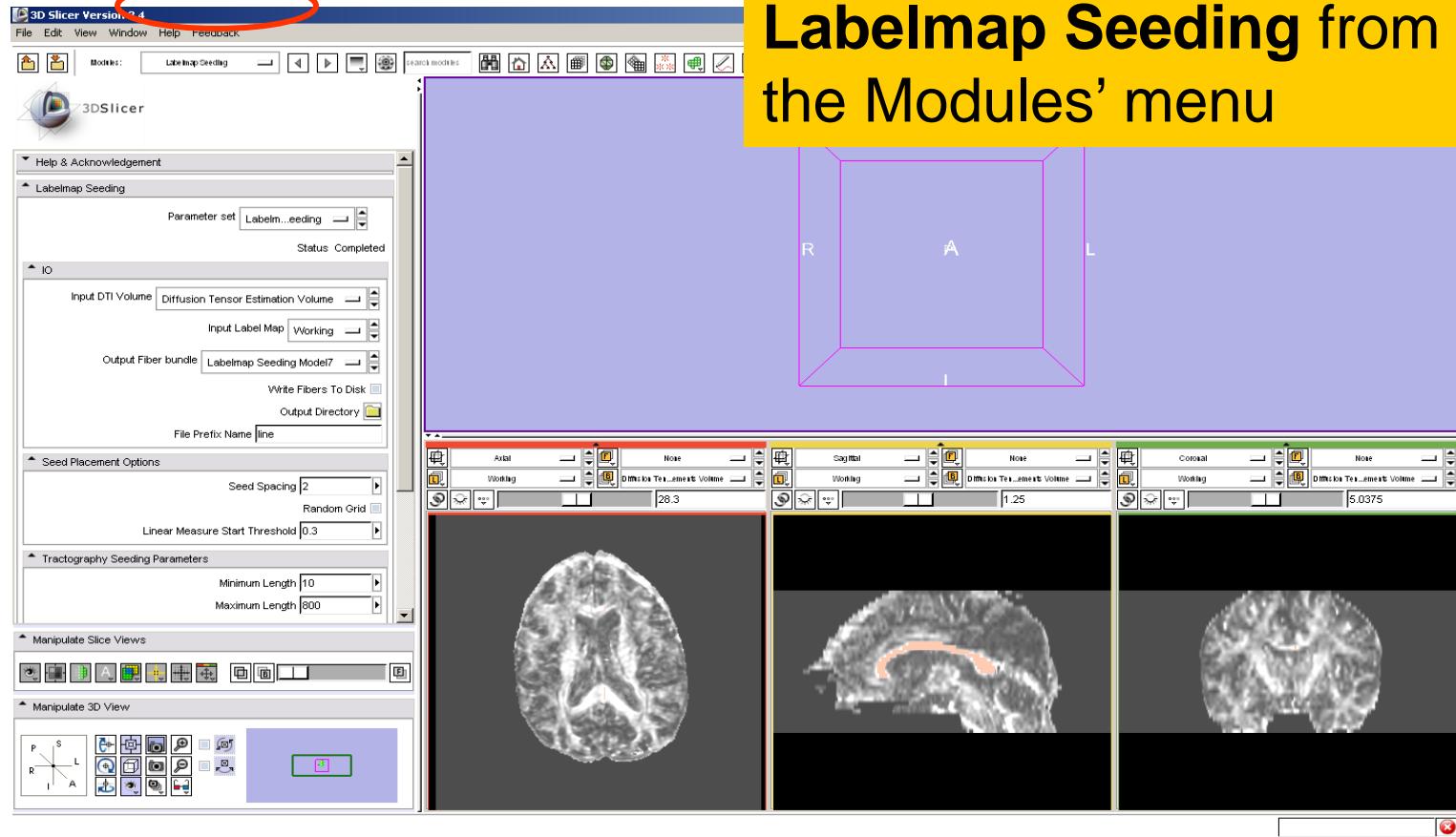


LabelMap Generation



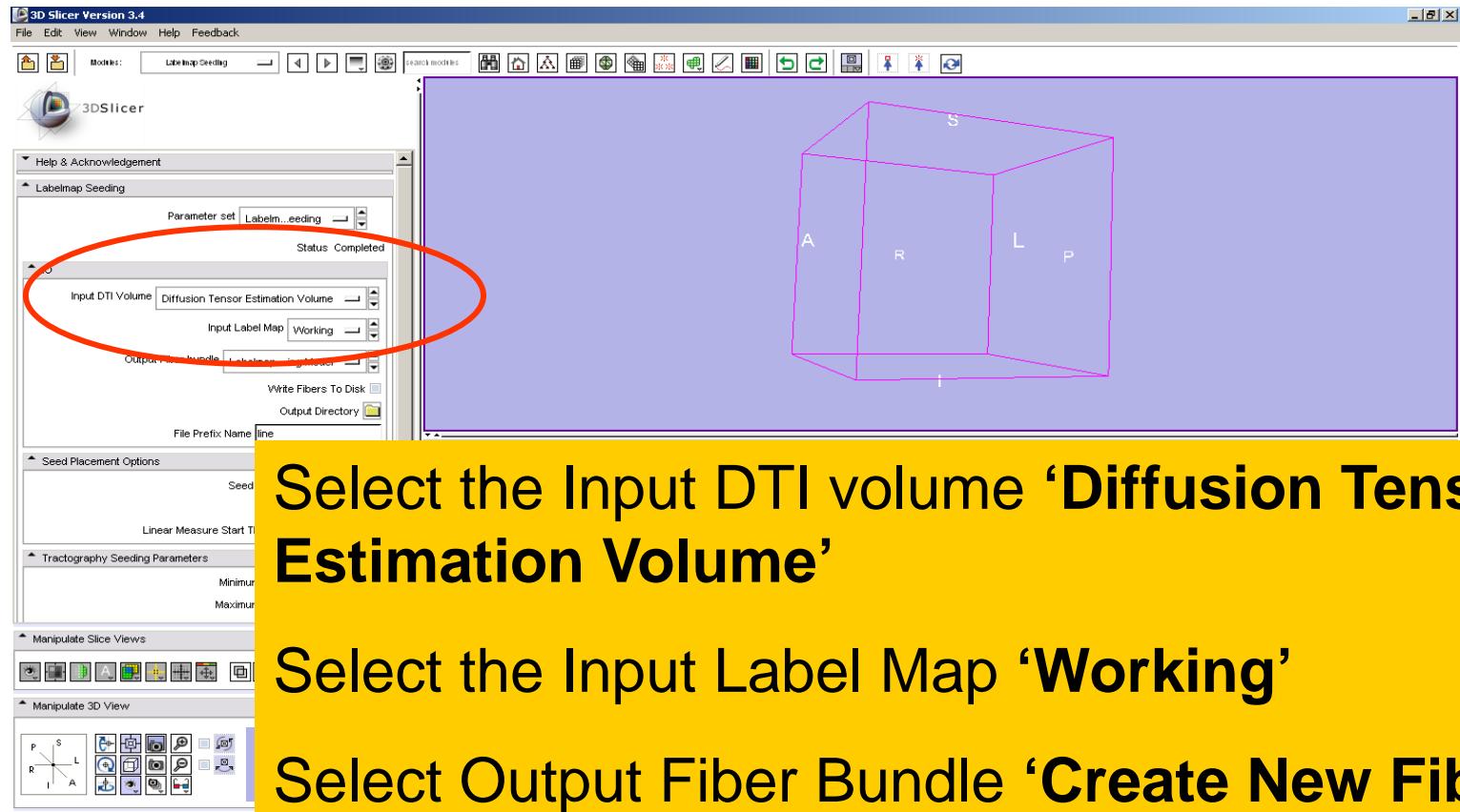


LabelMap Seeding





LabelMap Seeding





LabelMap Seeding

Set the Seed Spacing to **2 mm** and select the Stopping Mode **Fractional Anisotropy**

Use the default parameters for the minimum and maximum tract length, stopping value and stopping track curvature.

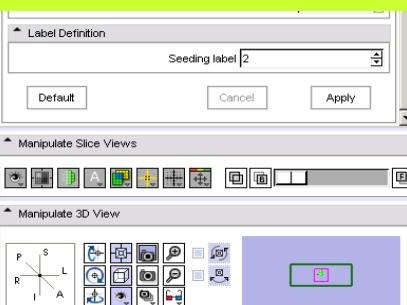
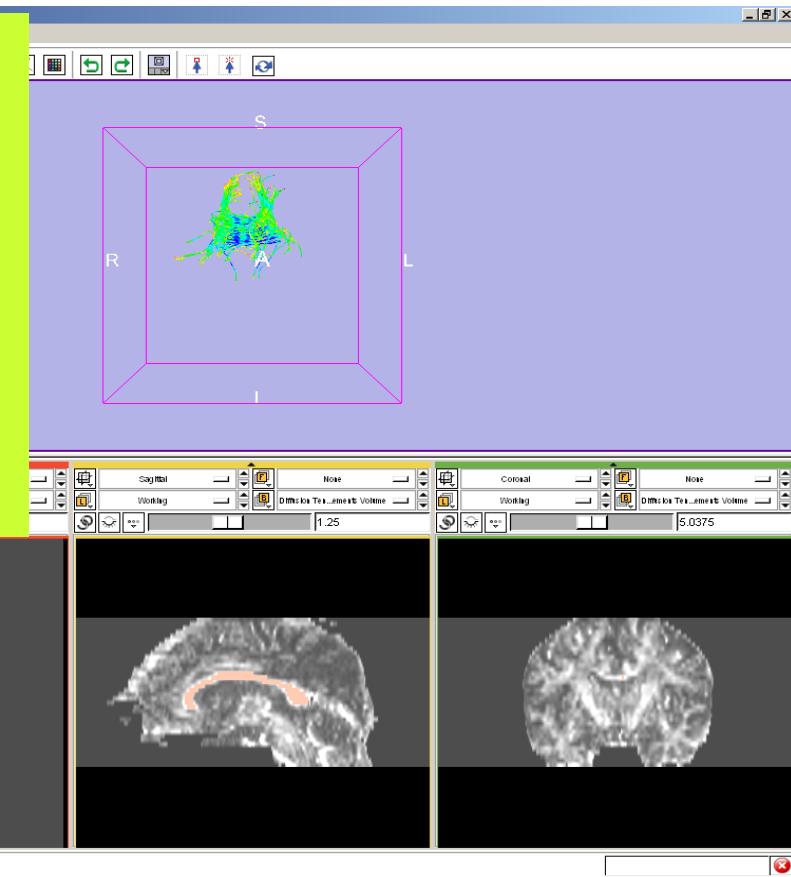
Set Seeding label to label 2, and click on **Apply**



LabelMap Seeding

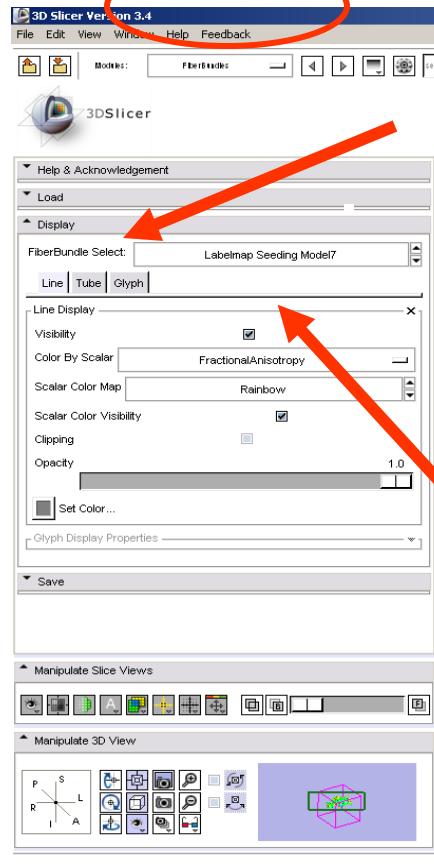
The tracts generated within the corpus callosum region appear in the 3DViewer.

The color map used represent the FA values along the tracts.

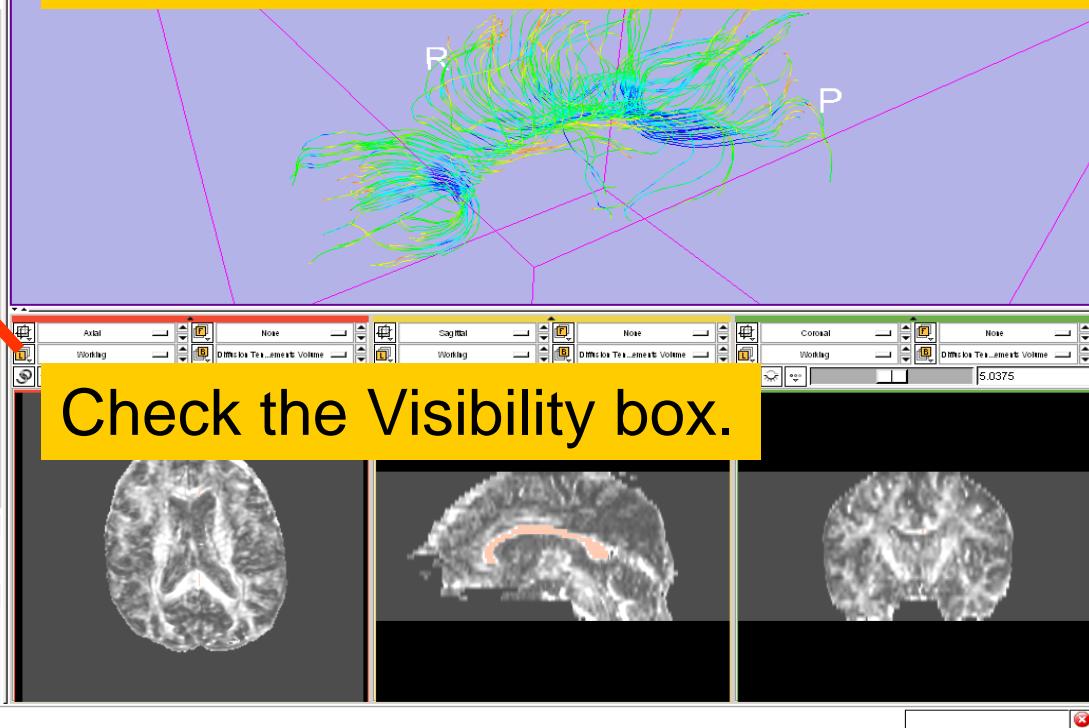




LabelMap Seeding

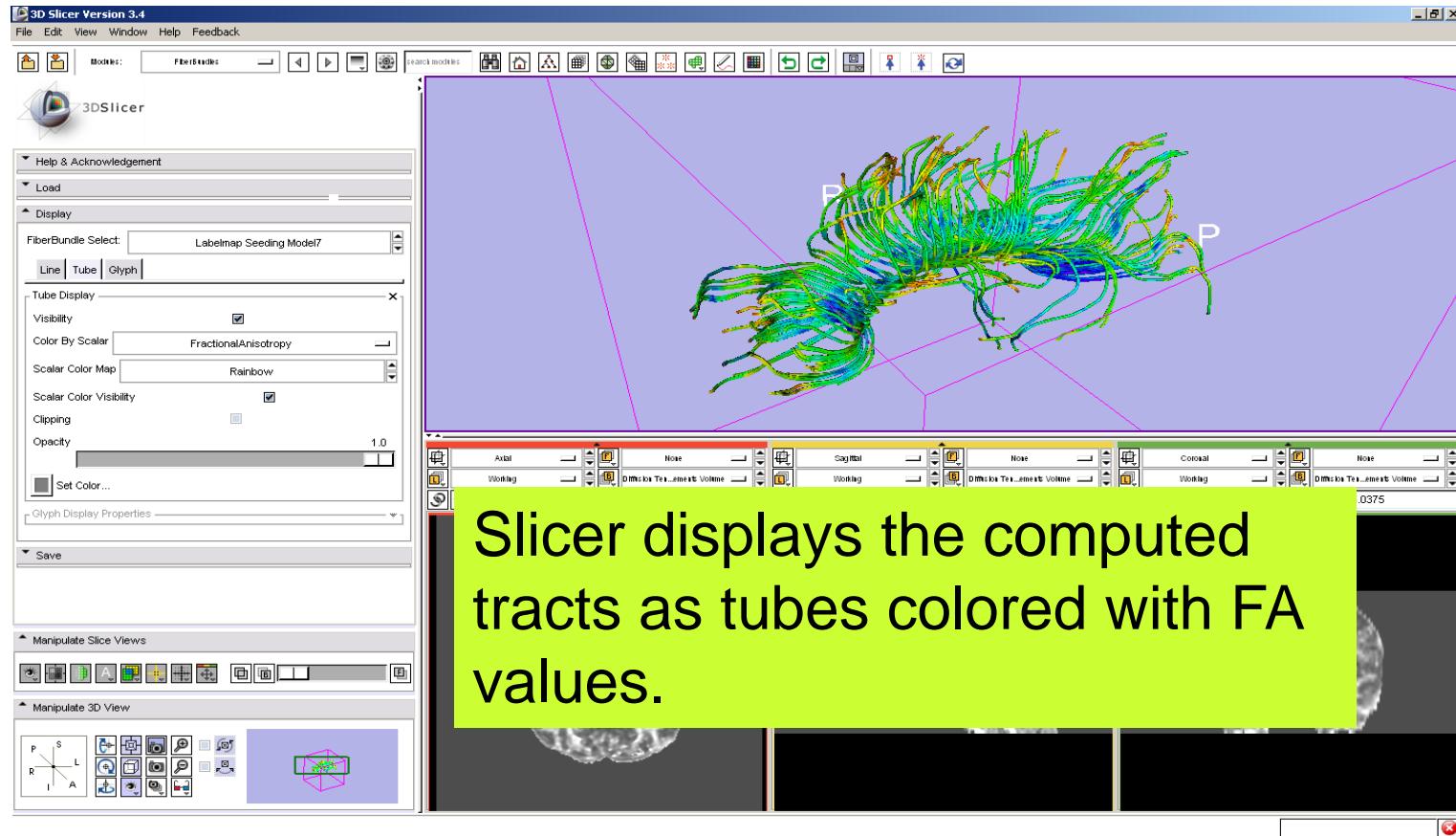


Select the module **FiberBundles**, and click on the tab **Tube** in the Display panel



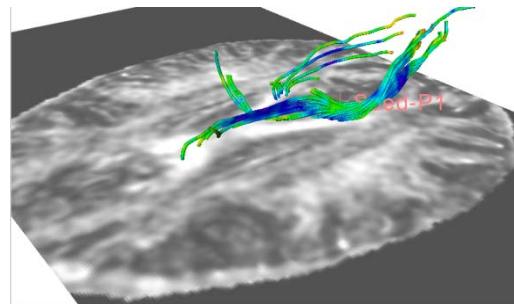
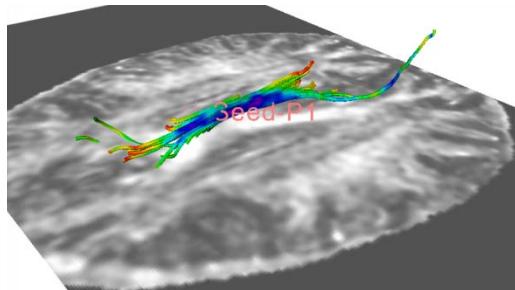
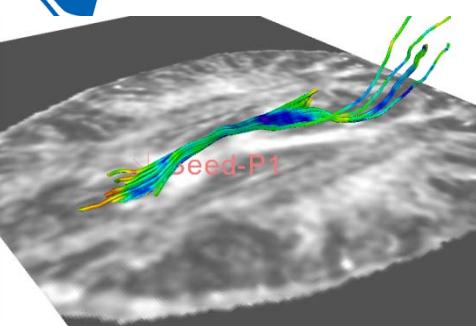


LabelMap Seeding



Part 4:

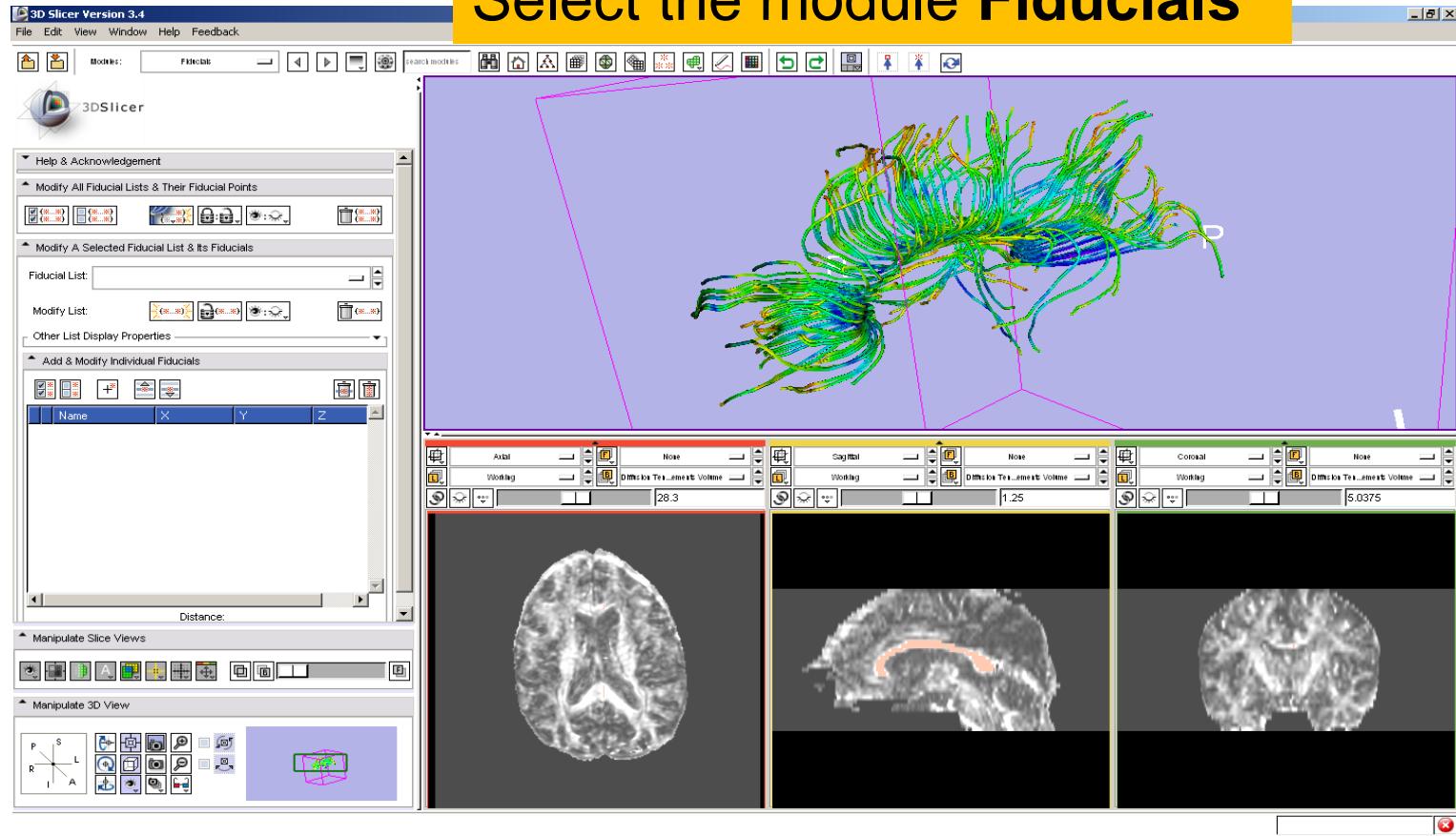
Tractography on-the-fly





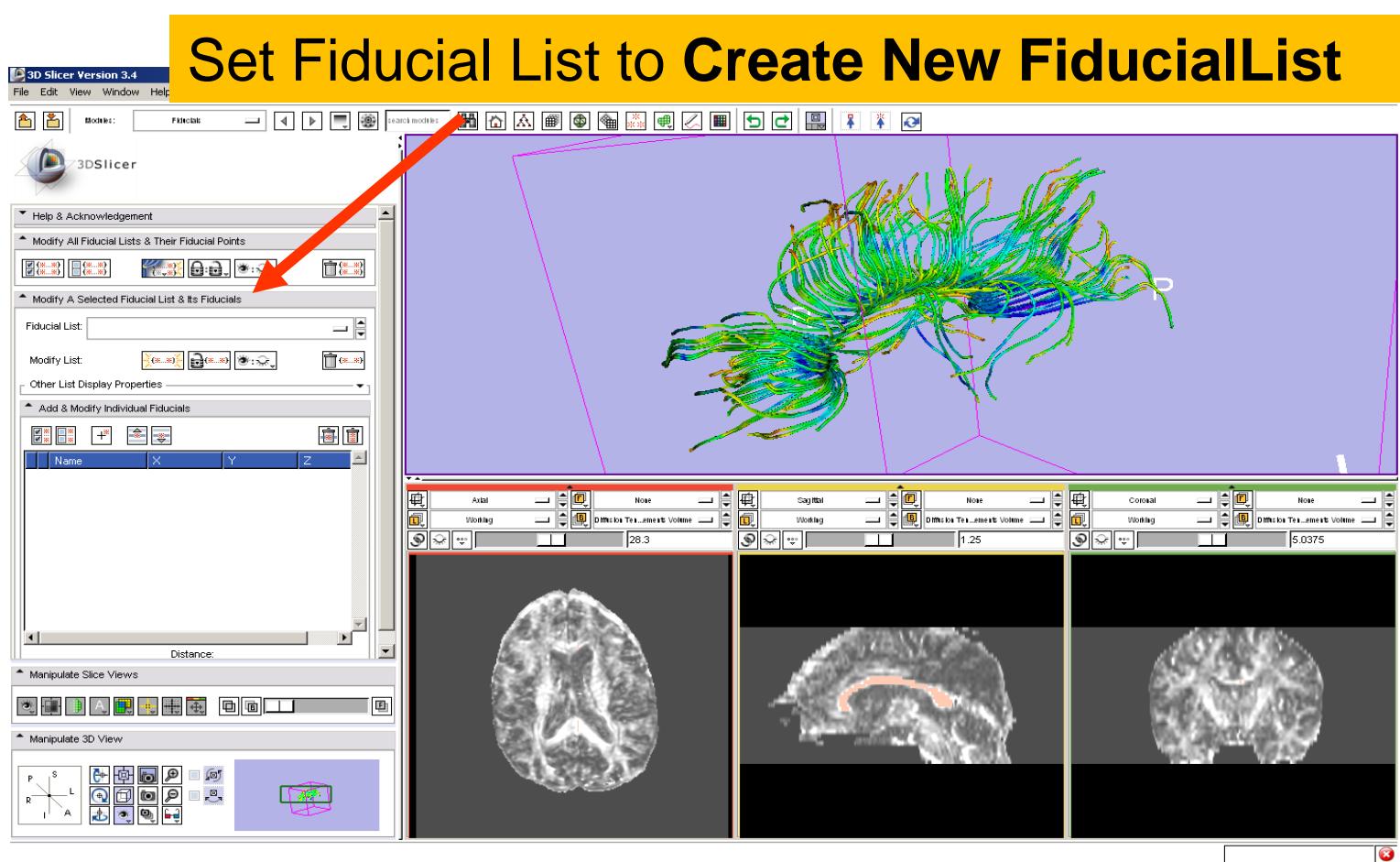
Fiducial Seeding

Select the module **Fiducials**



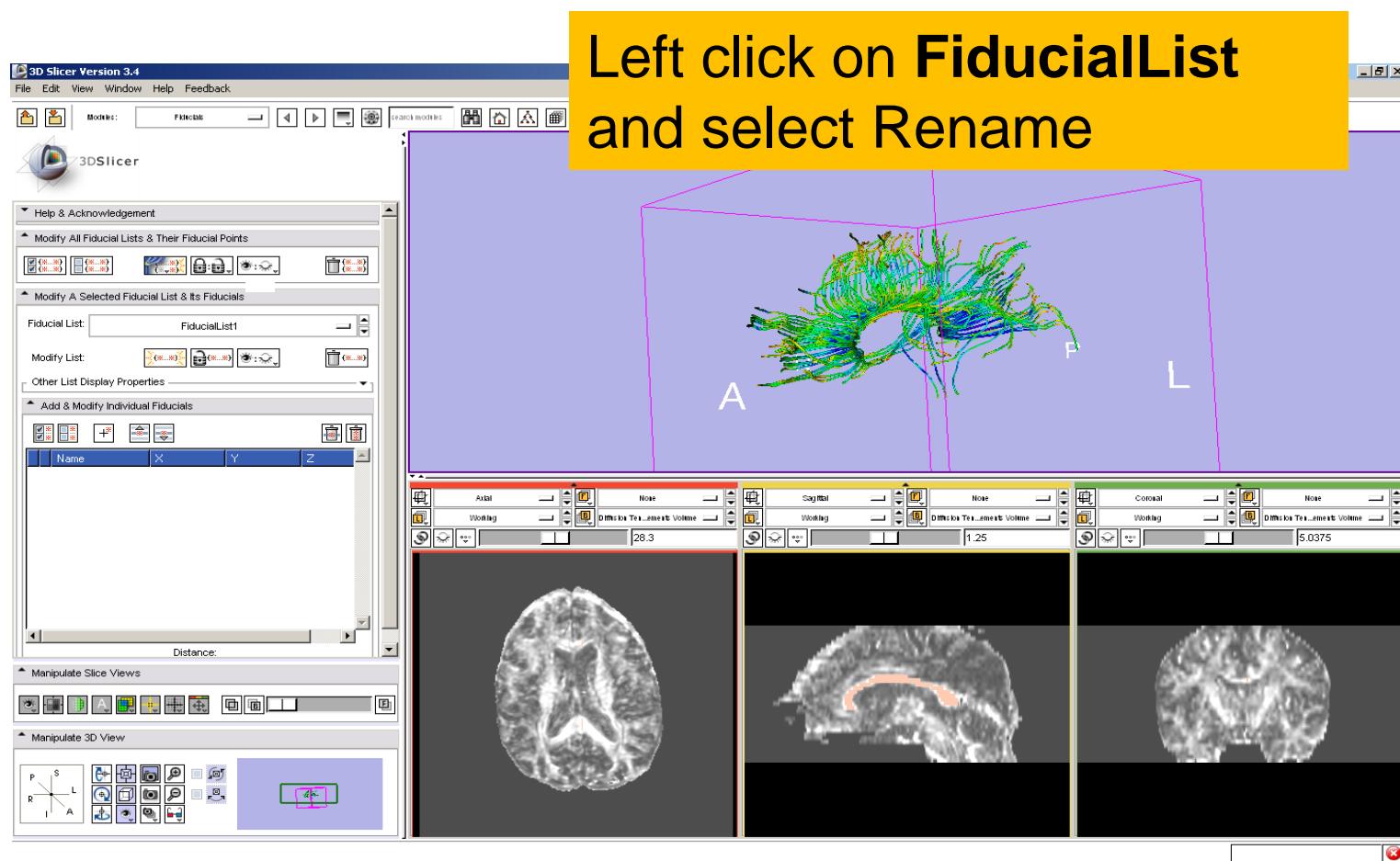


Fiducial Seeding





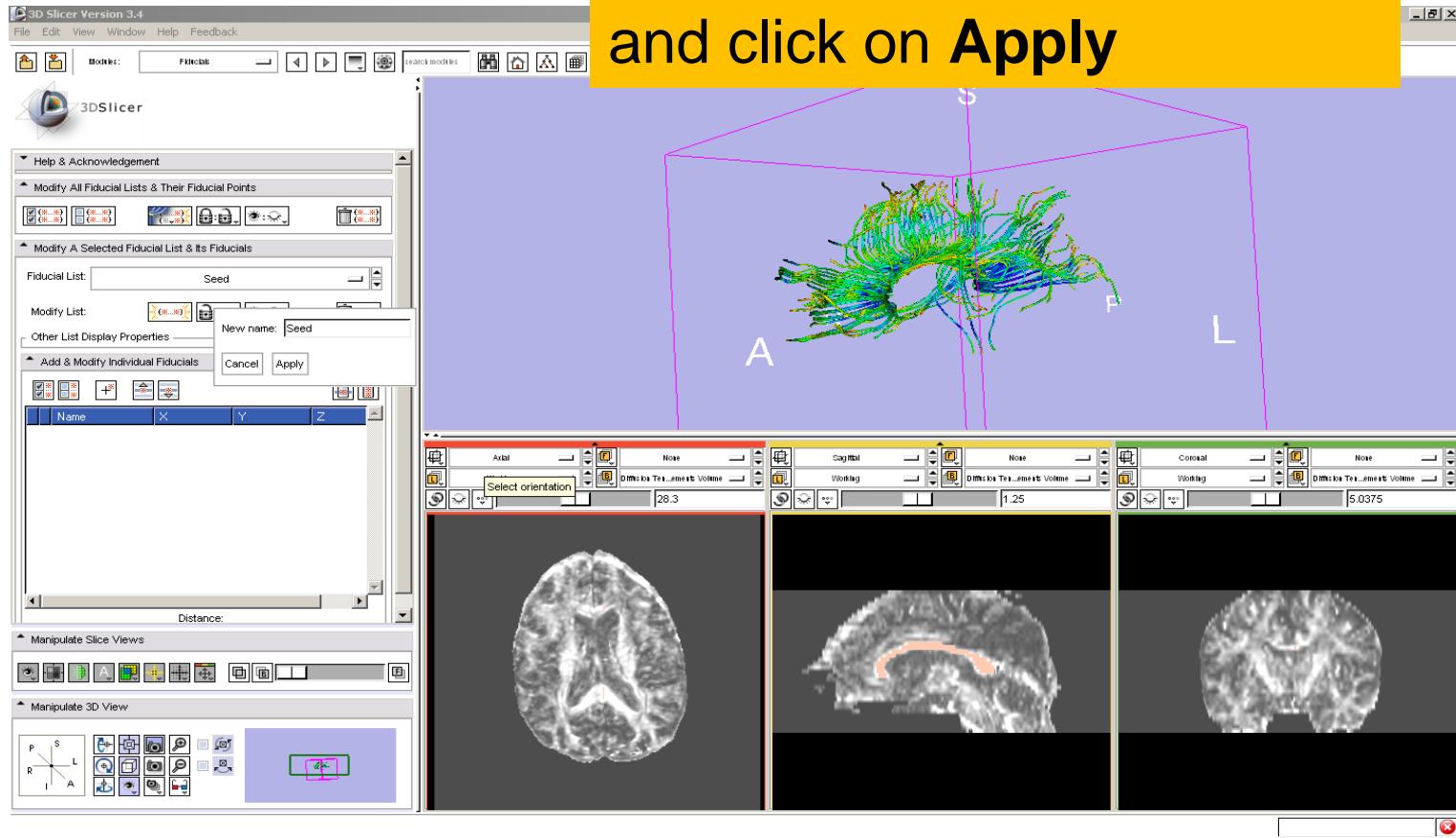
Fiducial Seeding





Fiducial Seeding

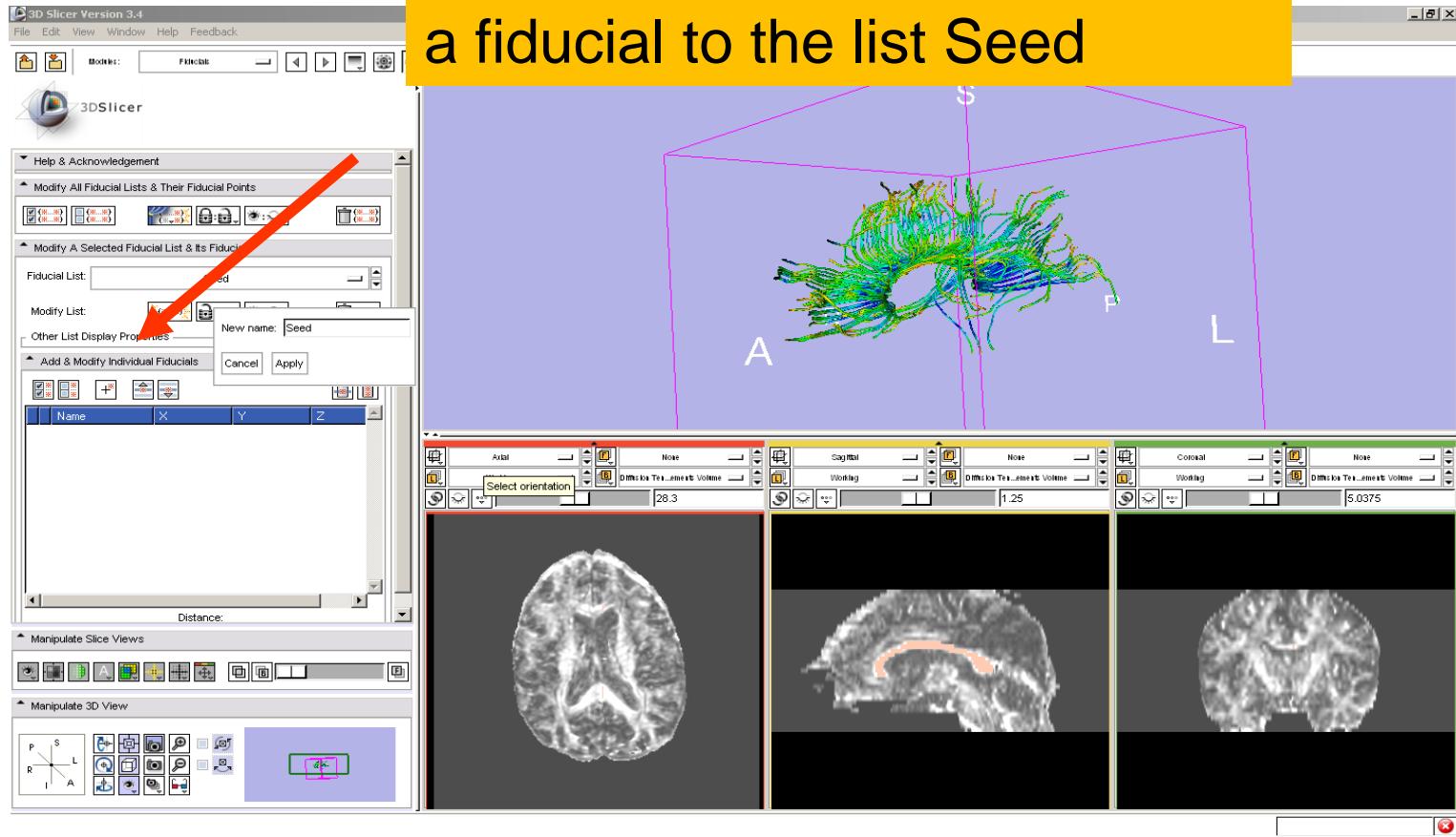
Enter the new name **Seed**
and click on **Apply**





Fiducial Seeding

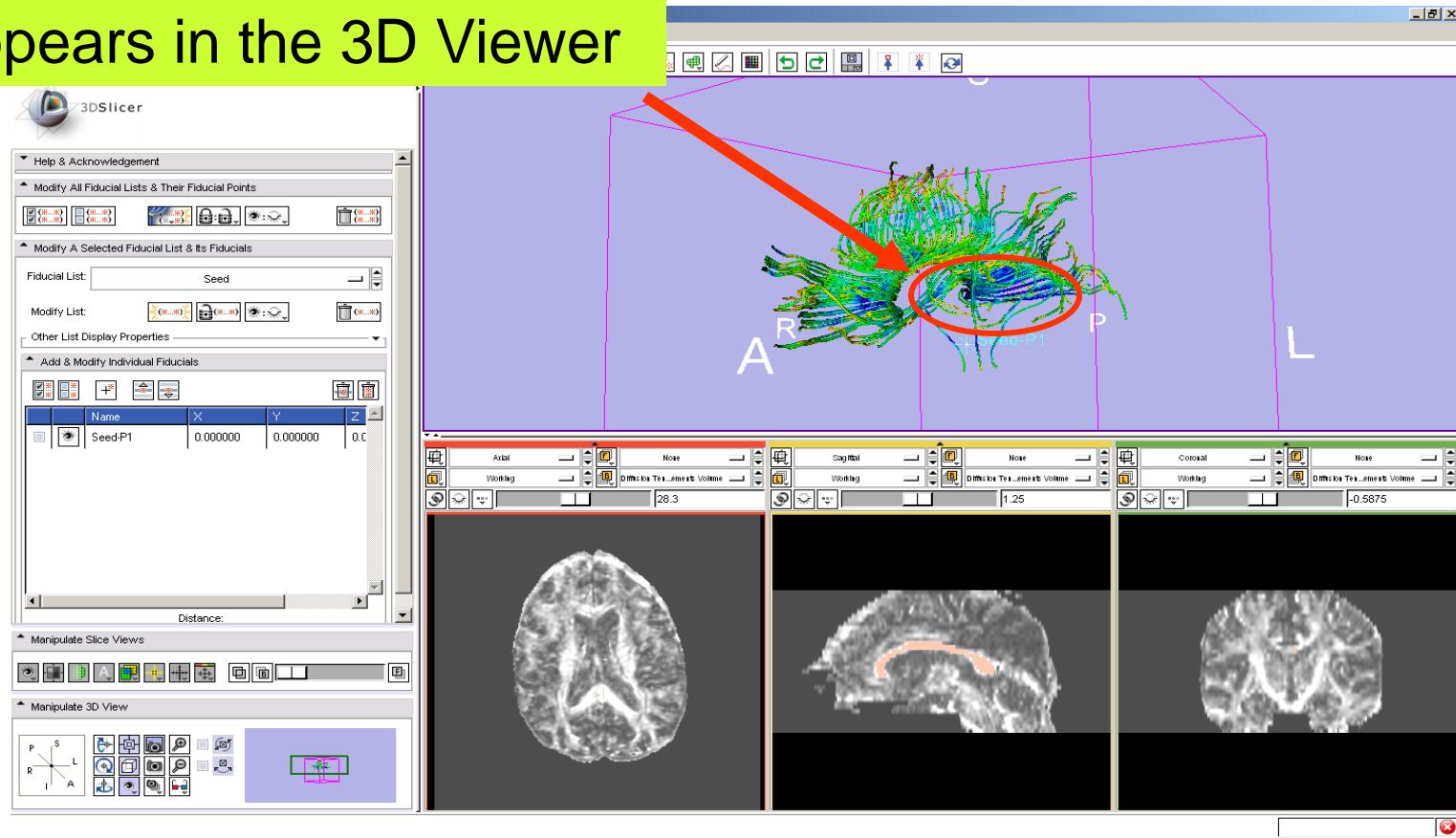
Click on the cross icon to add a fiducial to the list Seed





Fiducial Seeding

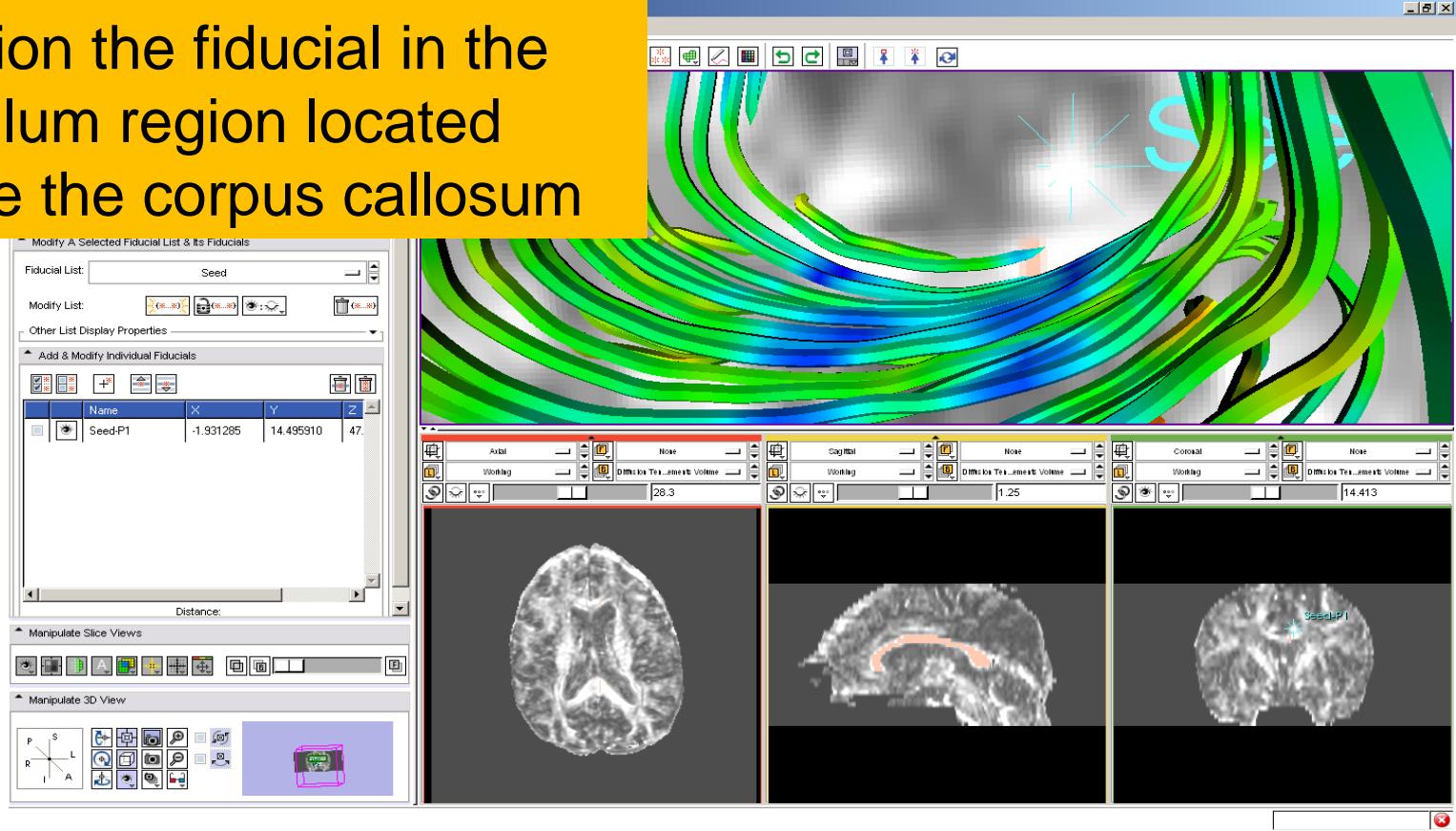
The fiducial **Seed-P1** appears in the 3D Viewer





Fiducial Seeding

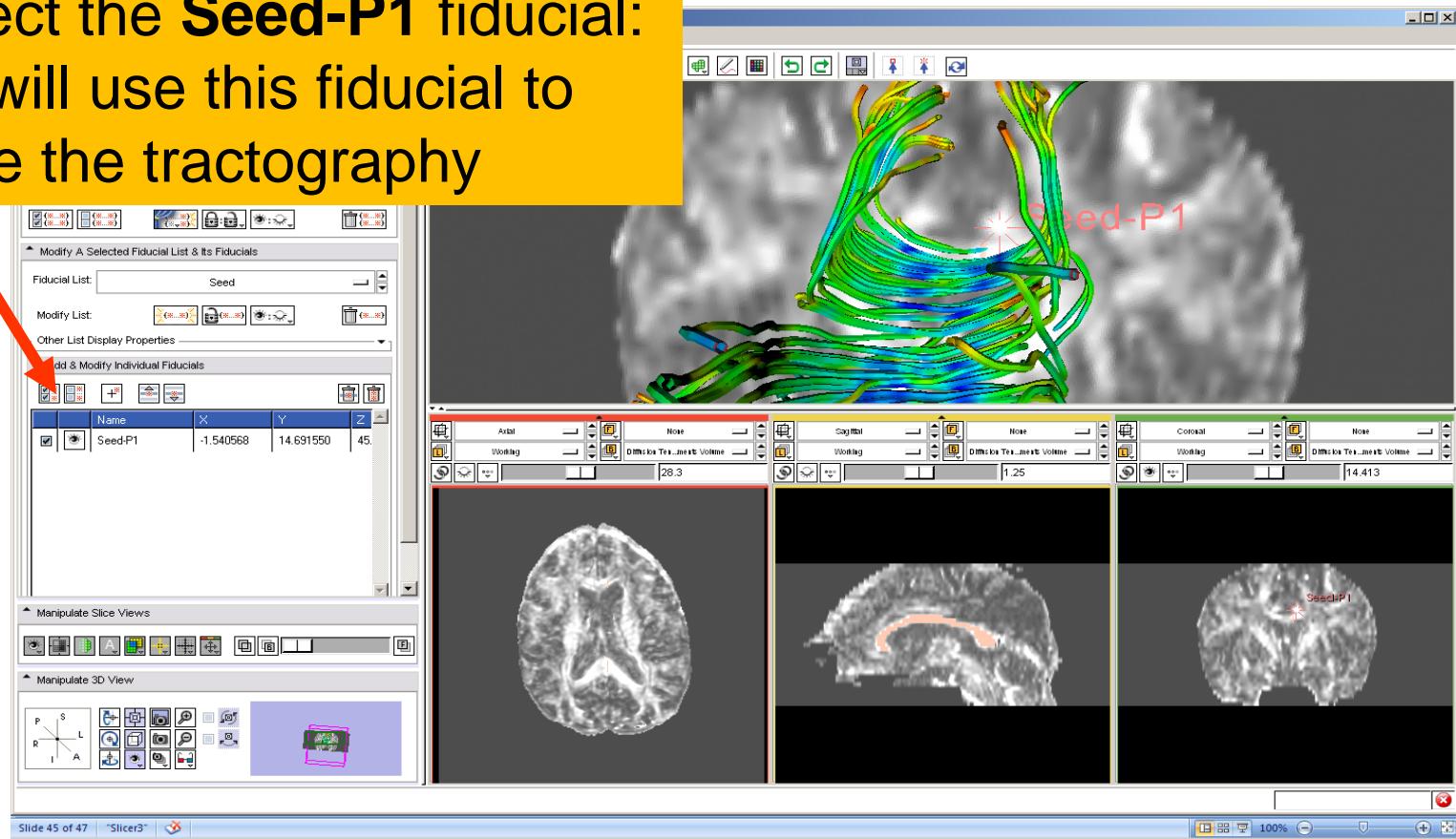
Position the fiducial in the cingulum region located above the corpus callosum





Fiducial Seeding

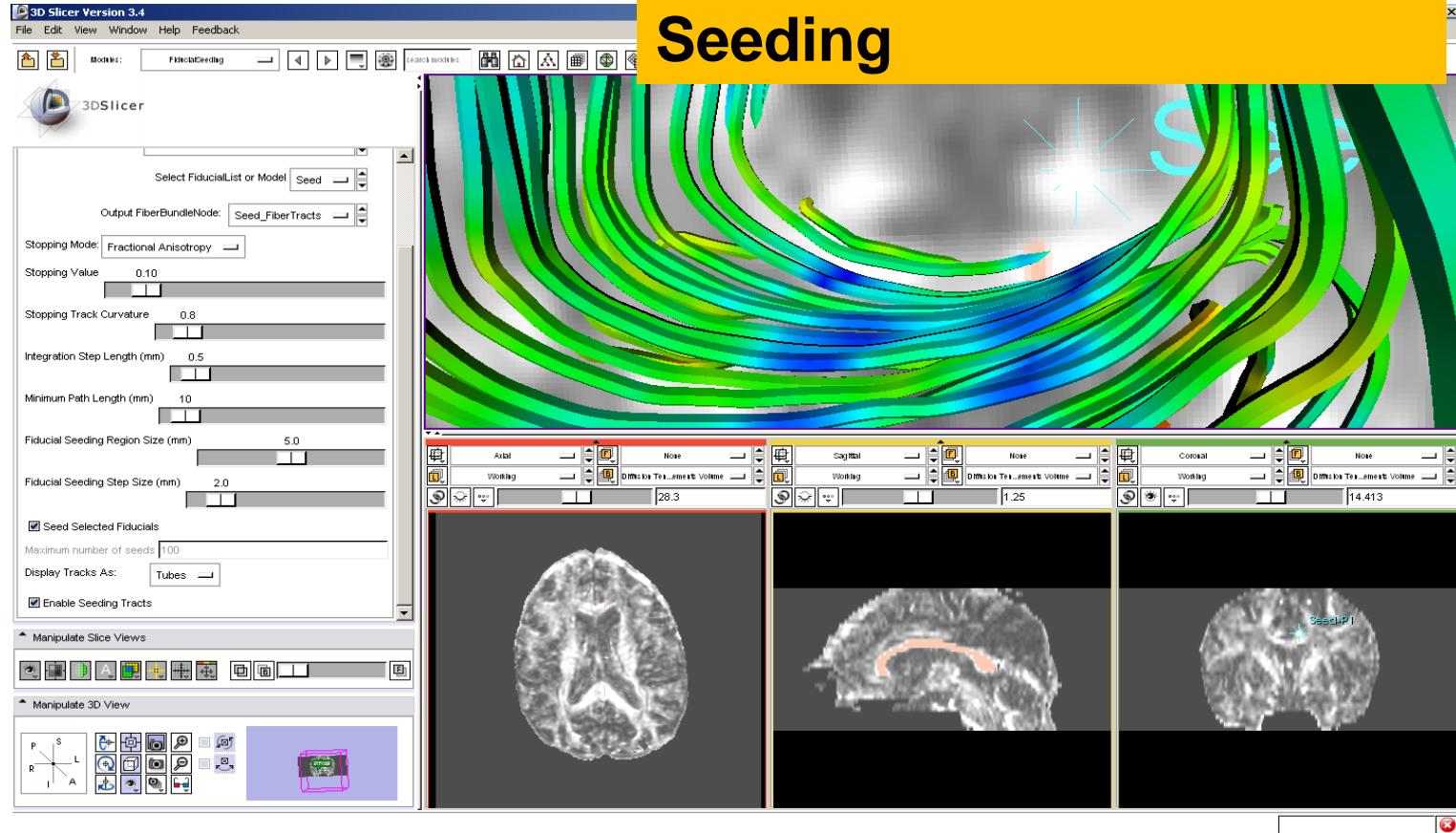
Select the **Seed-P1** fiducial:
we will use this fiducial to
drive the tractography





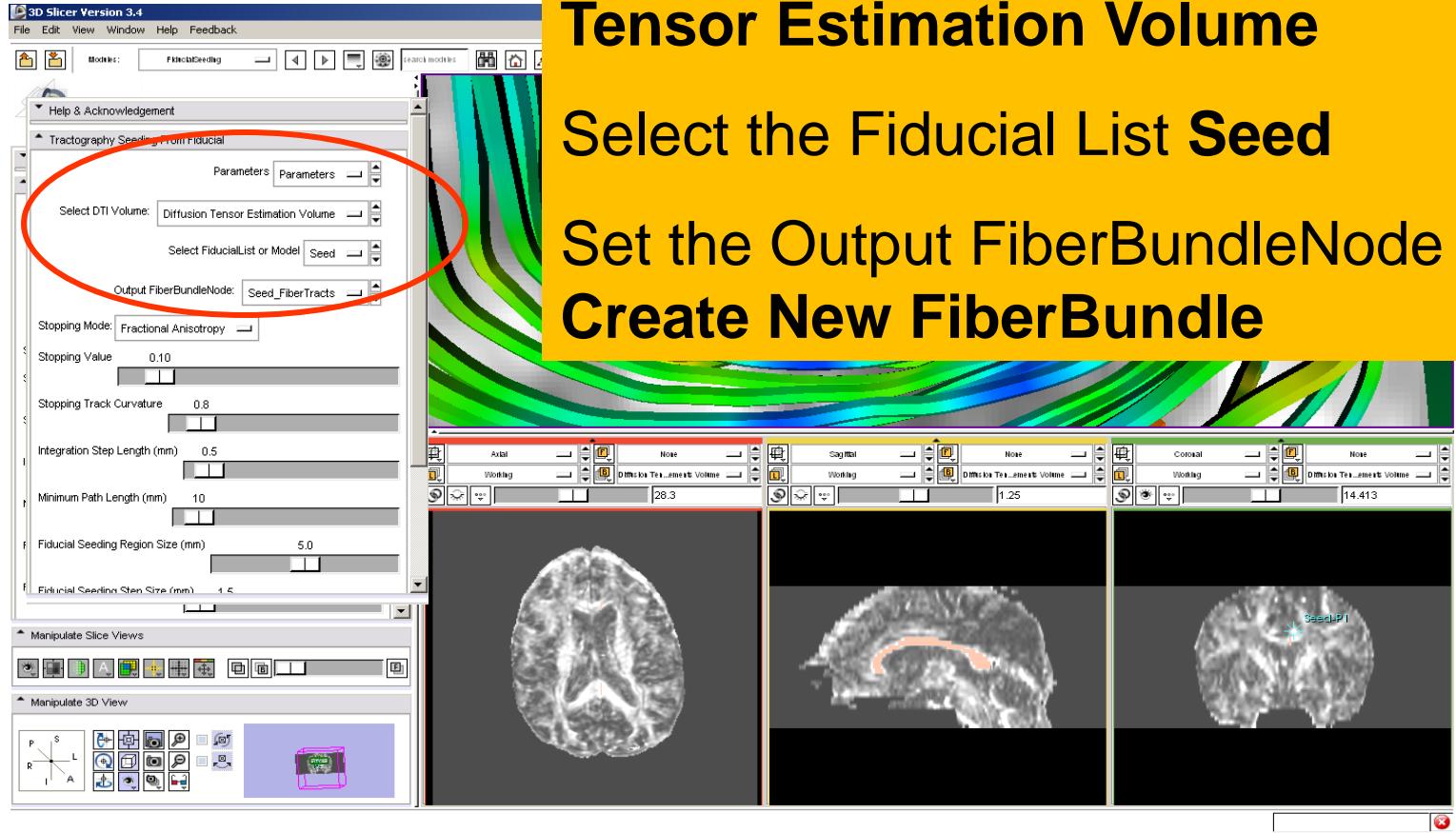
Fiducial Seeding

Select the module **Fiducial Seeding**





Fiducial Seeding



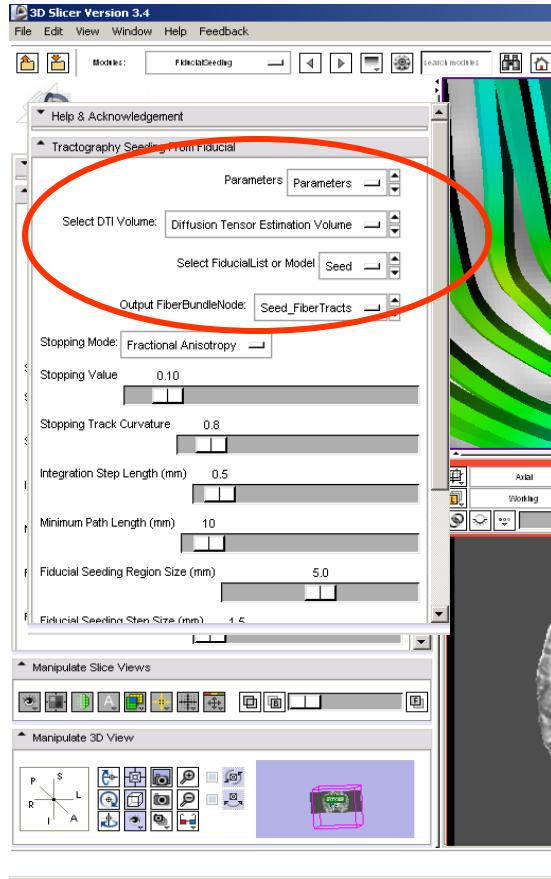
Set the DTI Volume to **Diffusion Tensor Estimation Volume**

Select the Fiducial List **Seed**

Set the Output FiberBundleNode to **Create New FiberBundle**



Fiducial Seeding



Set the Stopping Mode to Fractional Anisotropy and set the tractography parameters to the values that we used for the corpus callosum:

Stopping Value: 0.1

Stopping Track Curvature: 0.8

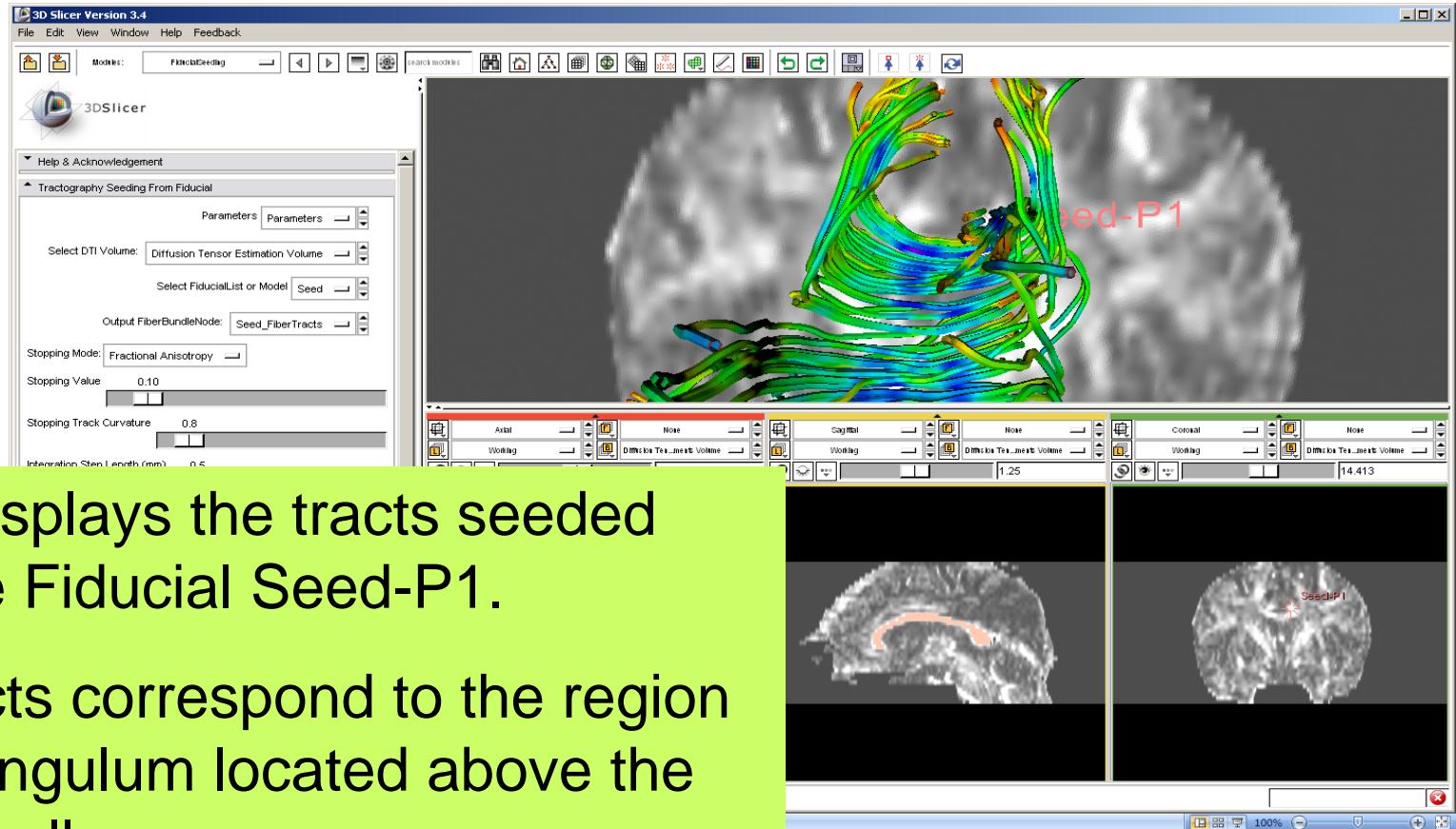
Step Length: 0.8 mm

Minimum Length: 10 mm

Fiducial Stepping Size: 1.5 mm

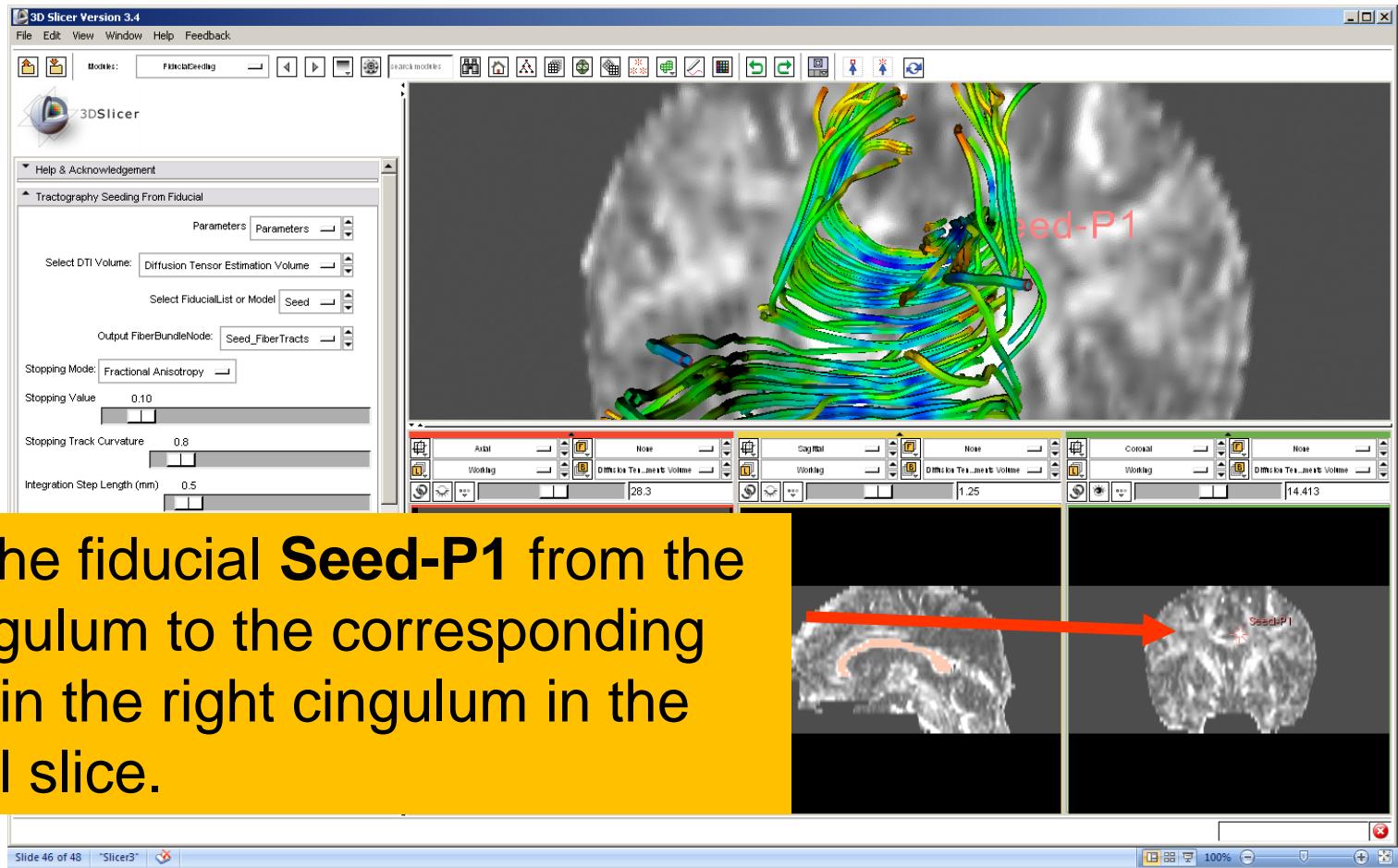


Fiducial Seeding





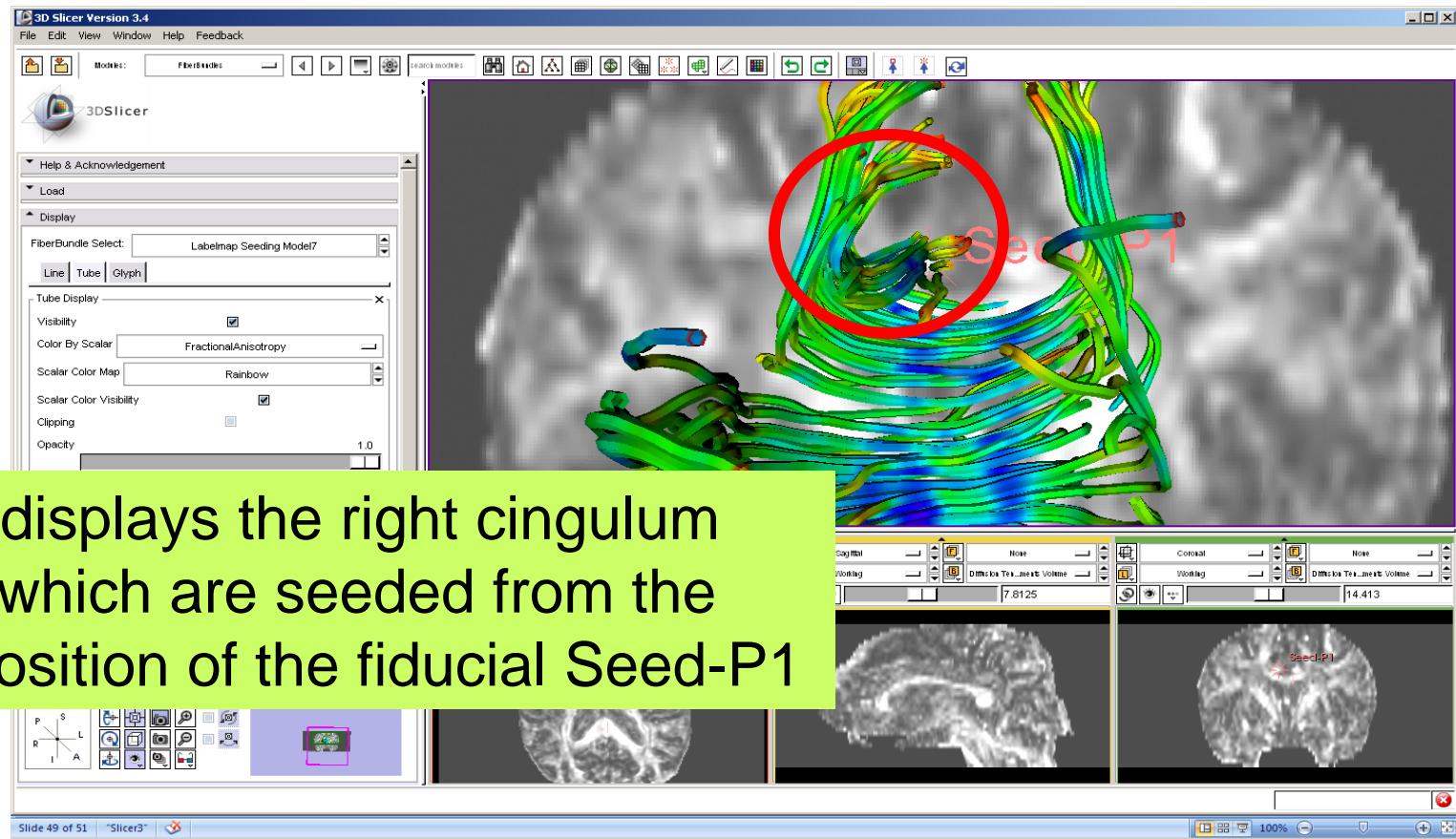
Fiducial Seeding



Move the fiducial **Seed-P1** from the left cingulum to the corresponding region in the right cingulum in the coronal slice.

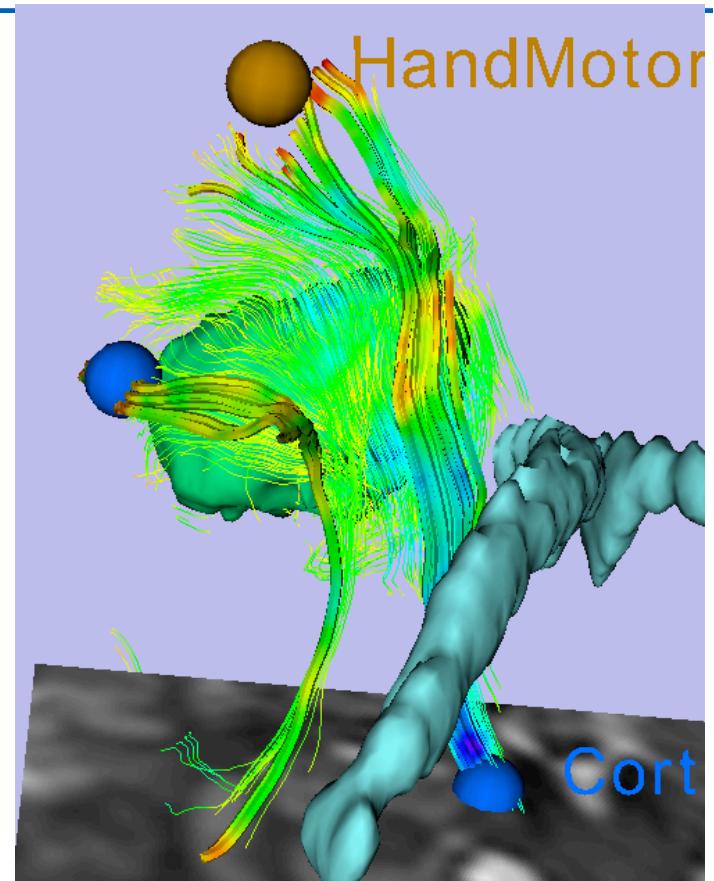
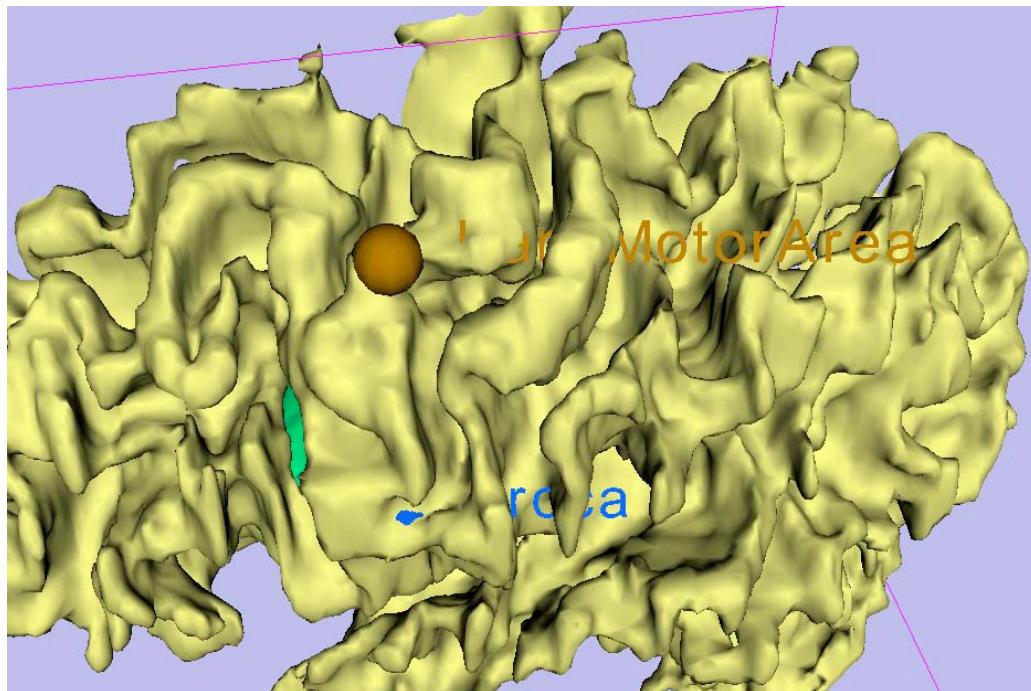


Fiducial Seeding

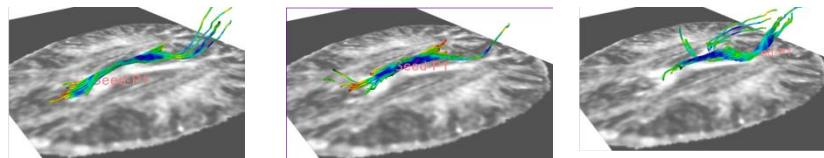
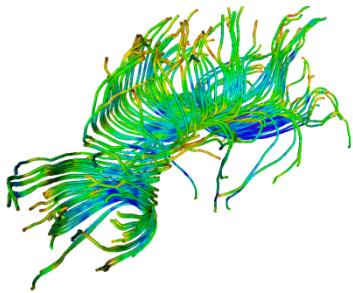
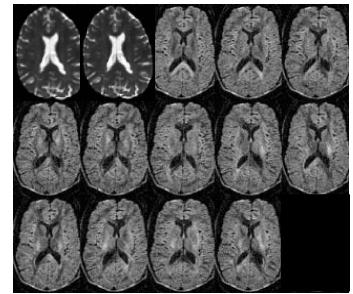




Neurosurgical Application



Courtesy of Alexandra Golby, MD, Peter Black, MD and Ron Kikinis, MD
Brigham and Women's Hospital, Boston, MA

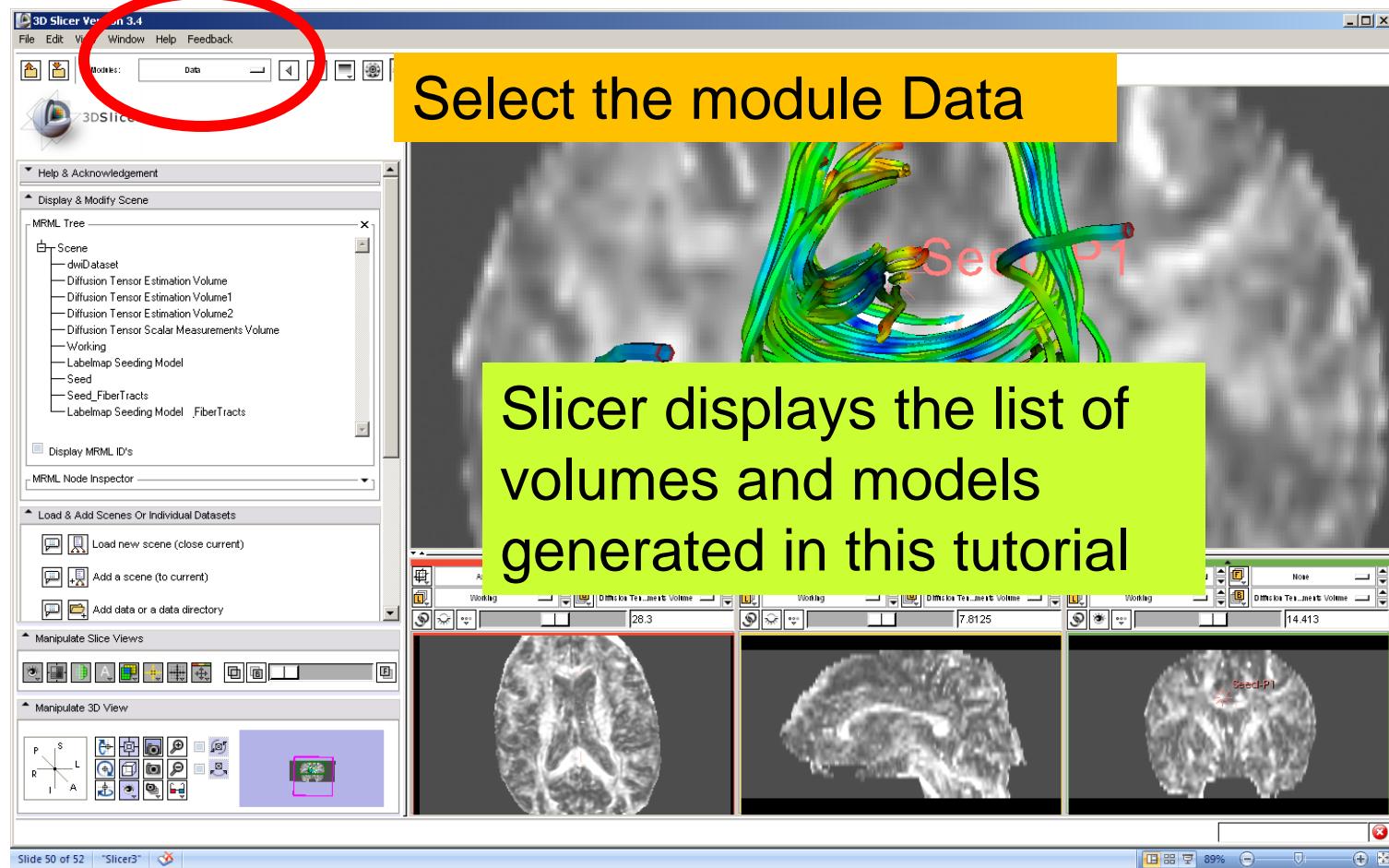


Part 5:

Saving a DTI Scene

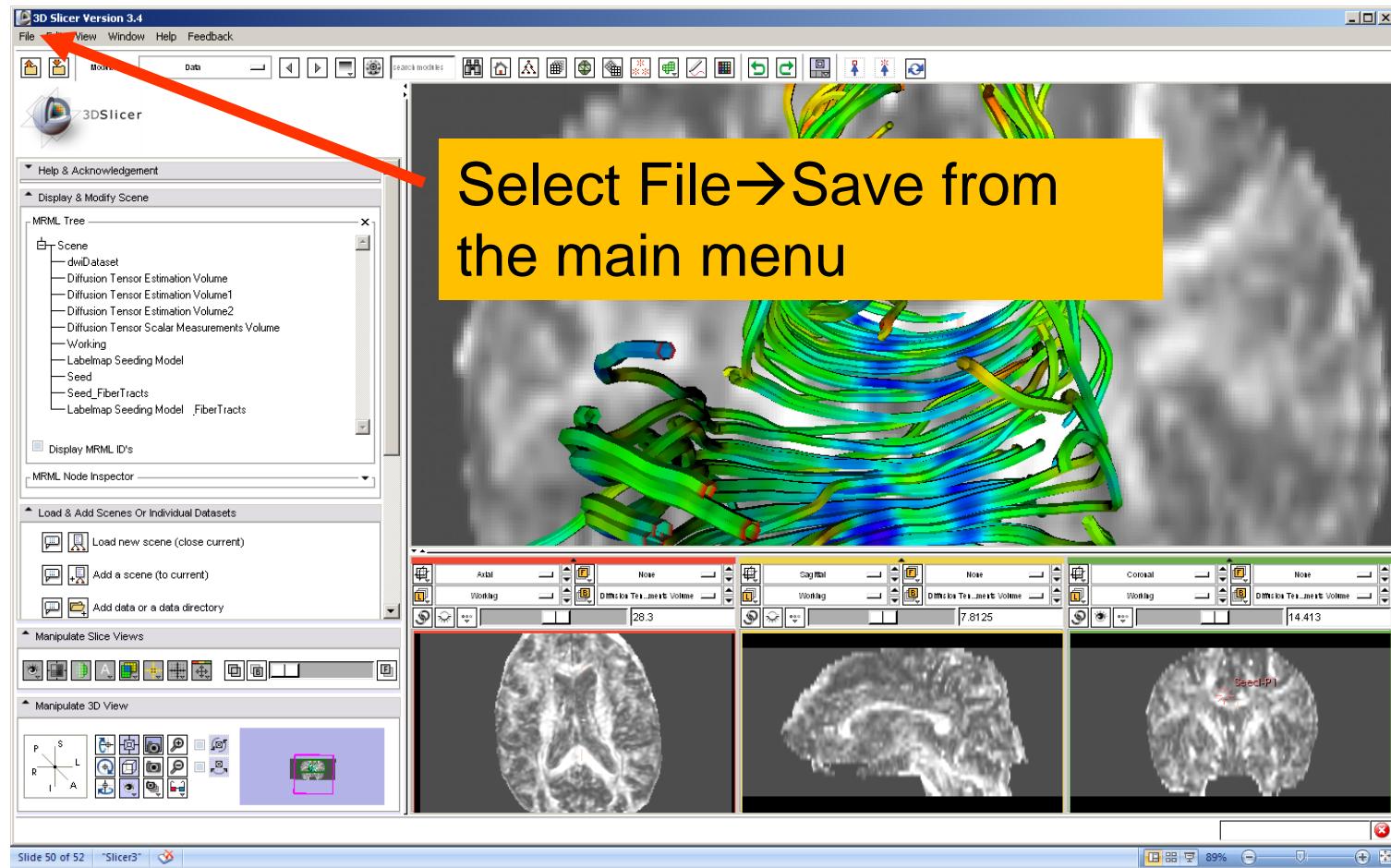


DTI Scene



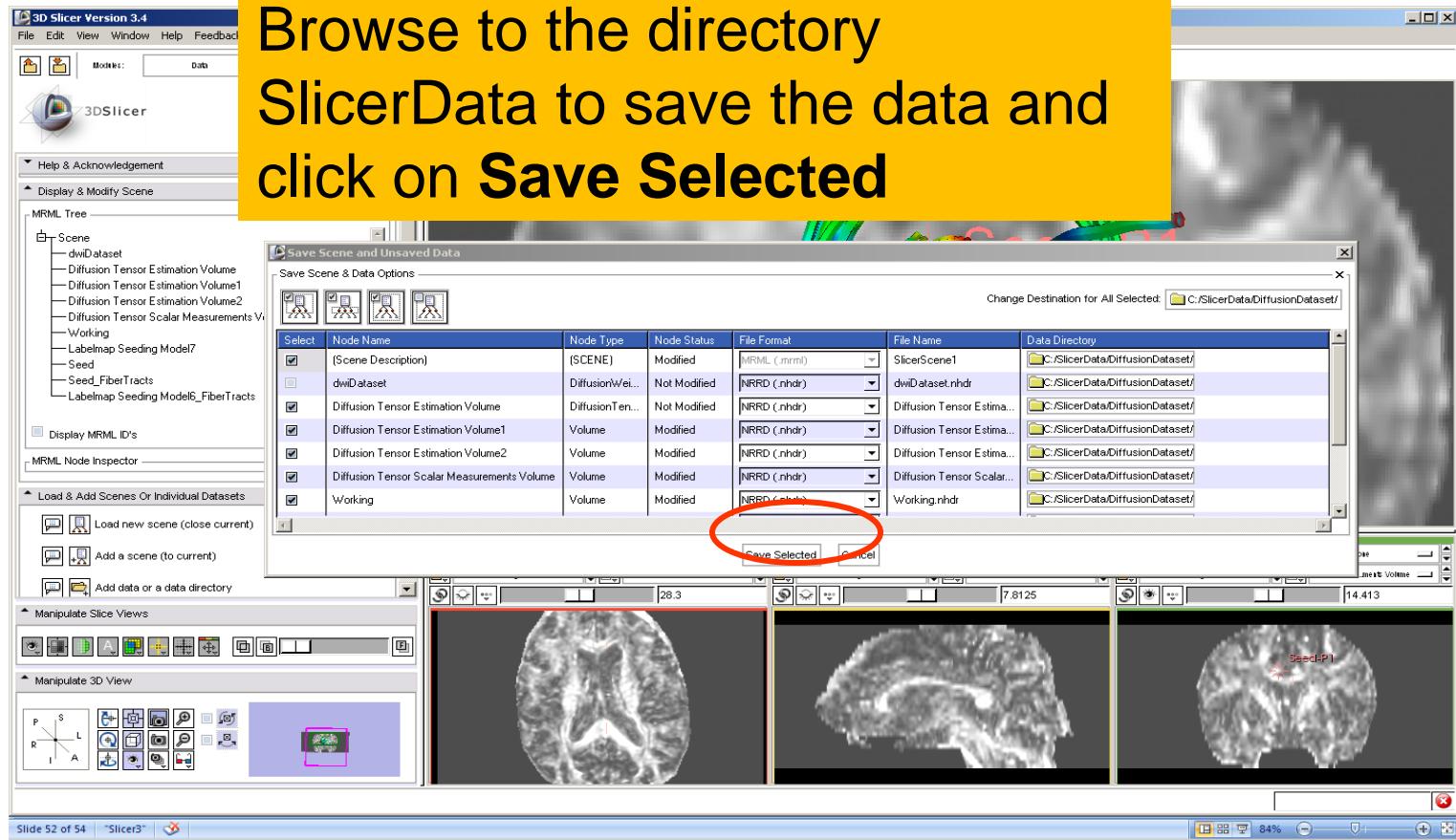


Saving a DTI Scene



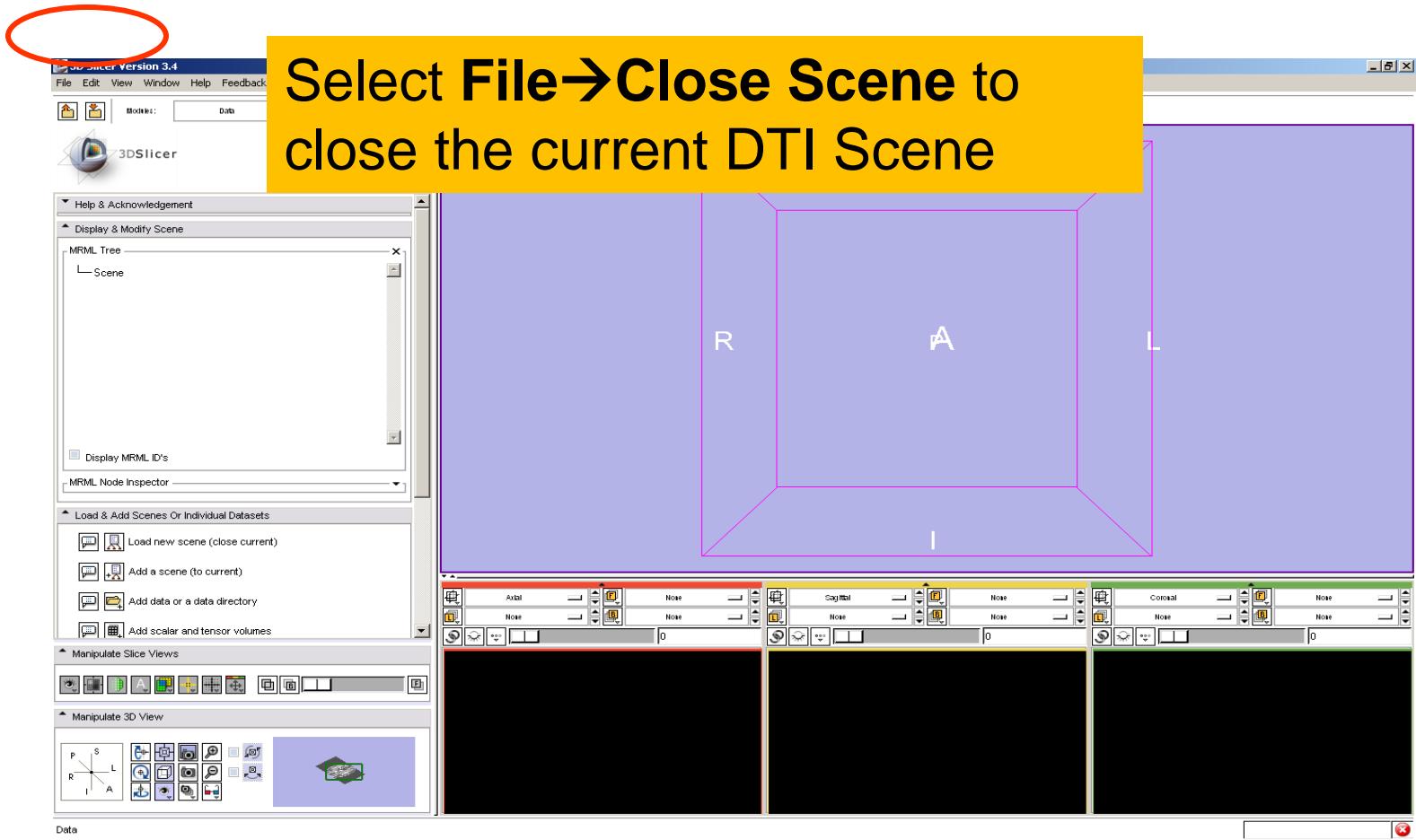


Saving a DTI Scene



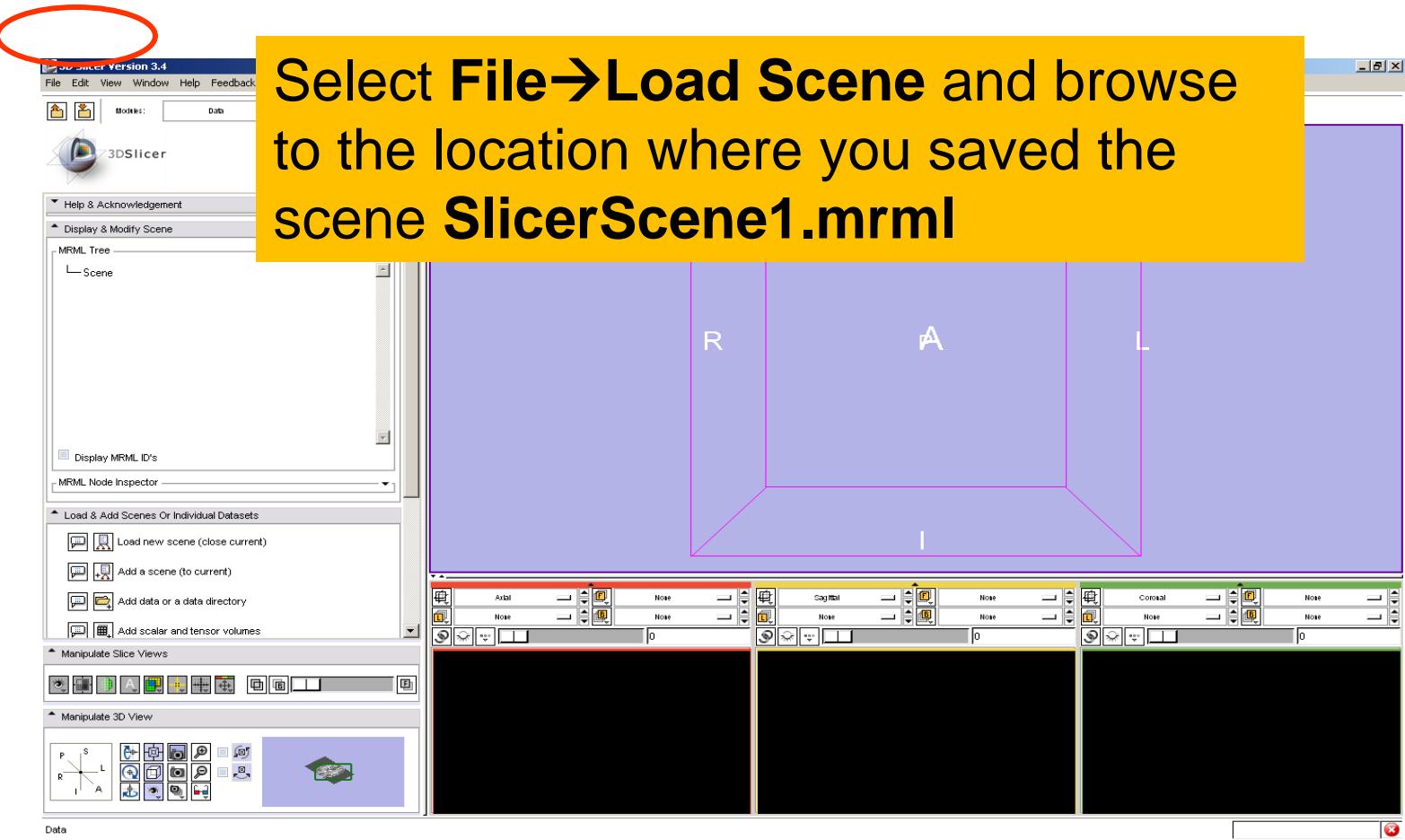


Saving a DTI Scene



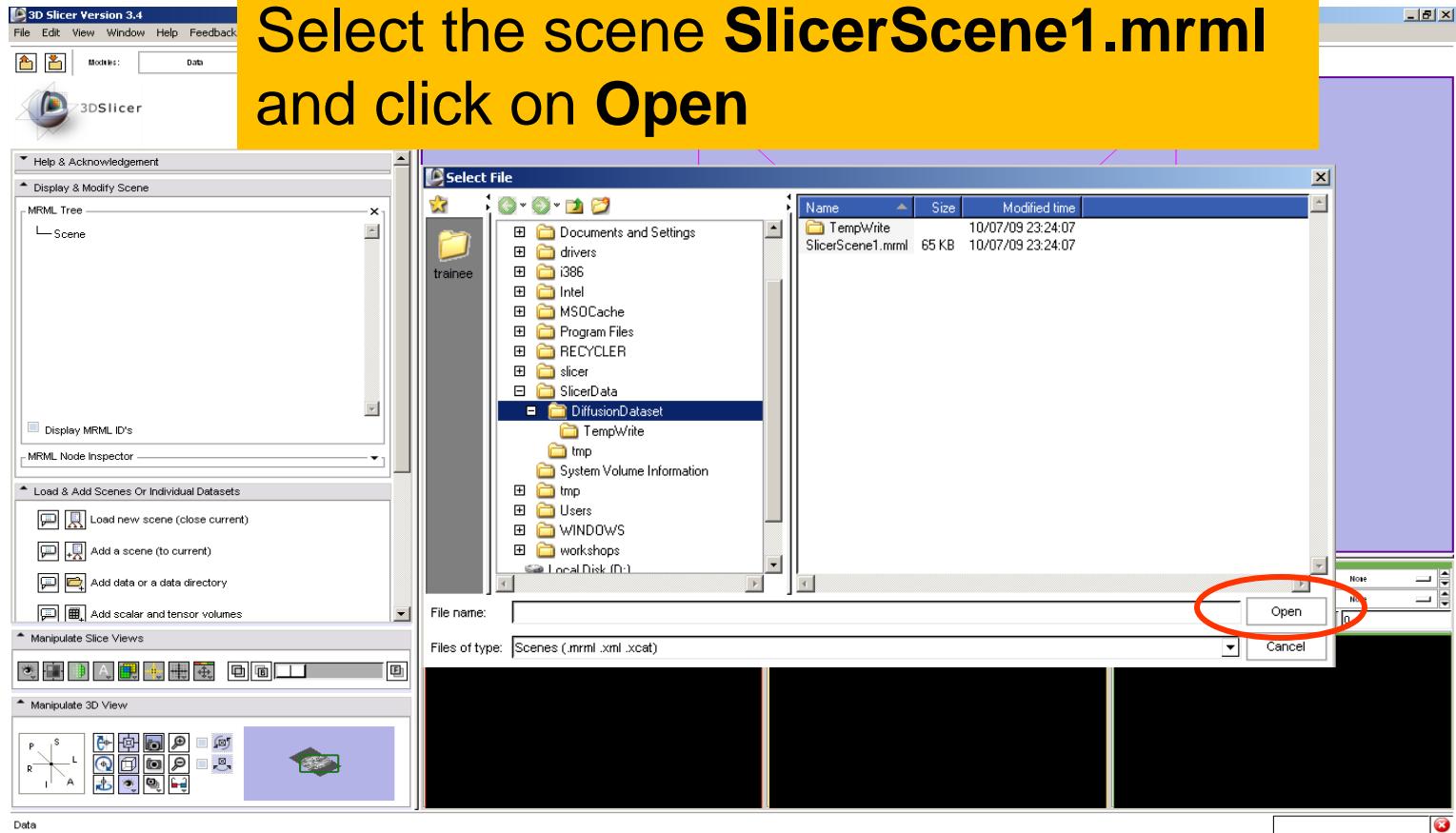


Loading a DTI Scene





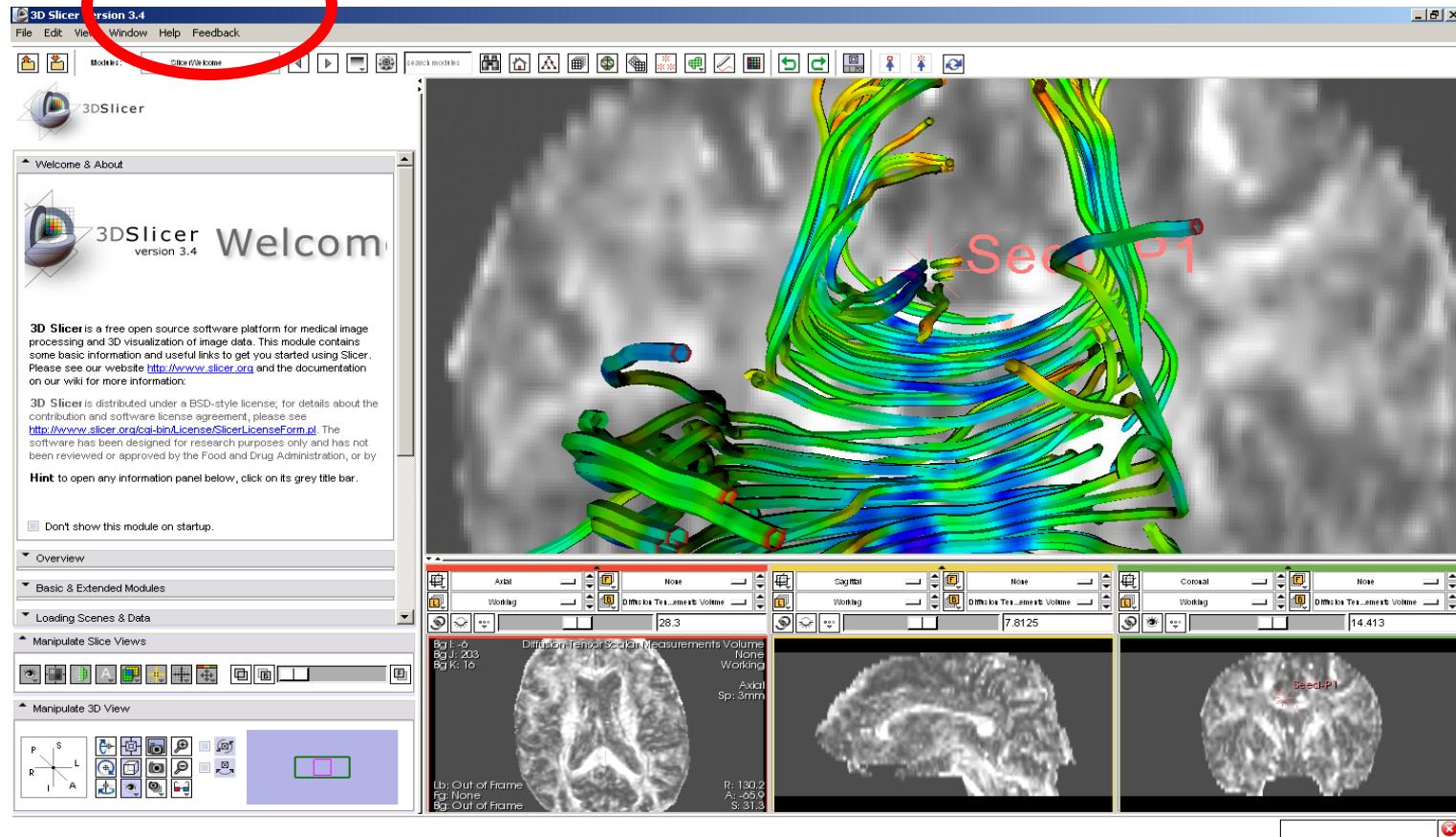
Loading a DTI Scene





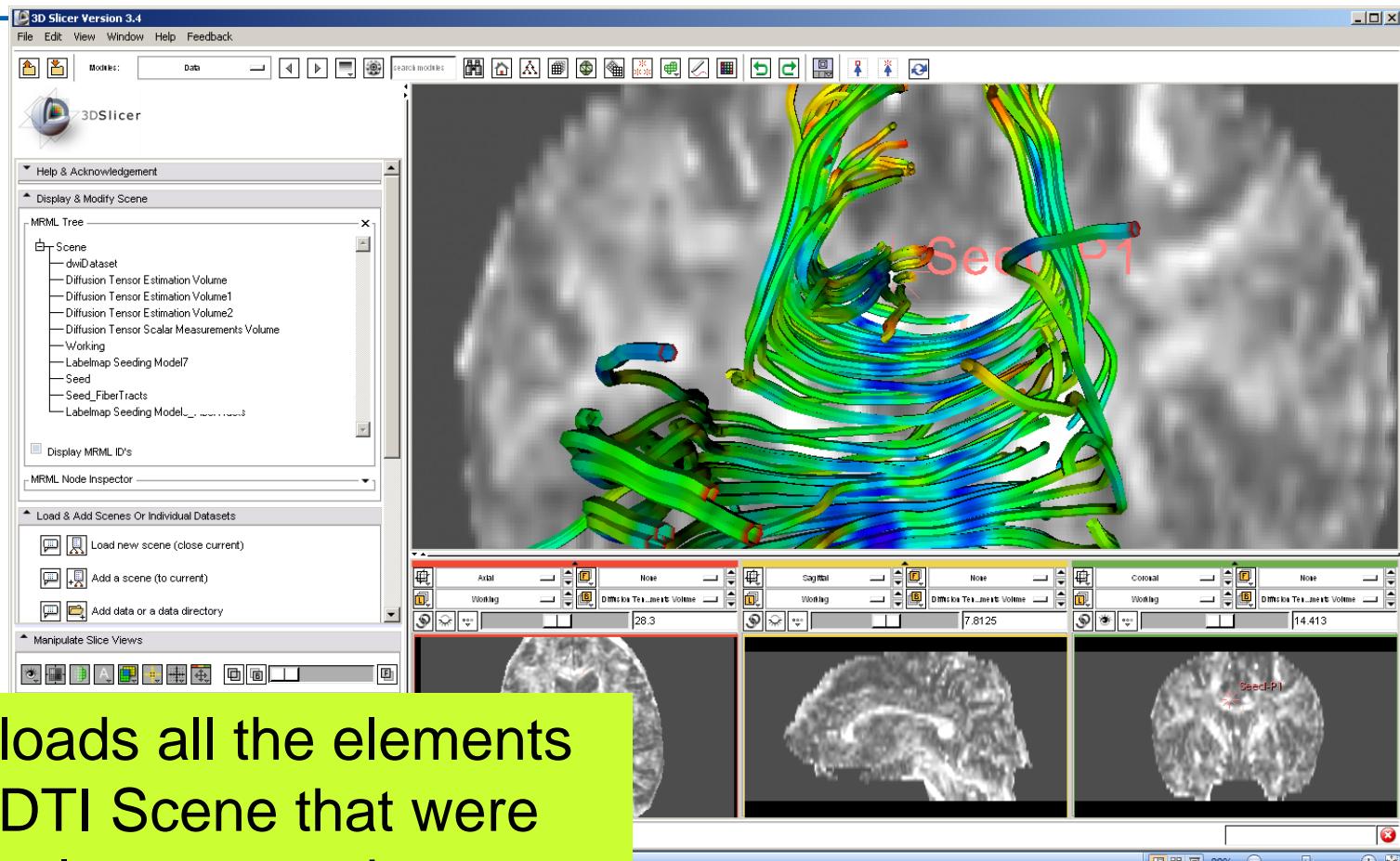
Loading a DTI Scene

Select the module Data





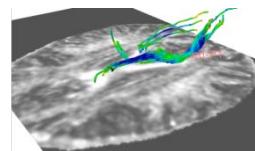
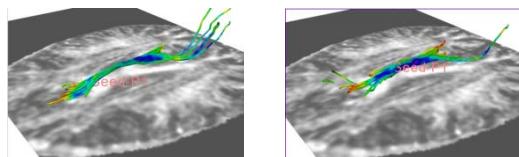
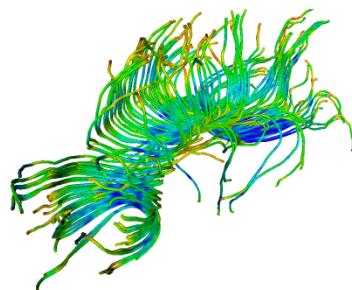
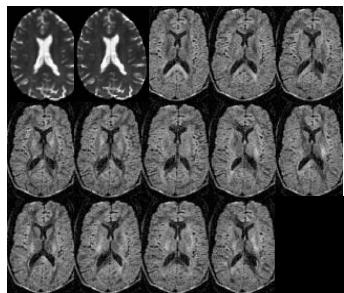
Loading a DTI Scene



Slicer loads all the elements of the DTI Scene that were previously computed.



Conclusion



This tutorial guided you through some of the **Diffusion MR** capabilities of the **Slicer3** software.

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